Biogeochemistry of the Bay of Bengal: Physical, chemical and primary productivity characteristics of the central and western Bay of Bengal during summer monsoon 2001

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Abstract

Reliable data on biological characteristics from the Bay of Bengal are elusive. In this paper, we present results on physics, chemistry and biology simultaneously measured during the summer monsoon, 2001 from open ocean and coastal areas of the region. It was characterised by cold core eddies and thermocline oscillations. However, these were capped by prevalent low saline upper regime and prevented surfacing of nutrients. The river plume effects were evident from the low salinity values observed in the surface layers of the upper bay, but this did not bring-in significant amount of nutrients. The chlorophyll *a* concentrations (10-20 mg m⁻²) and primary productivity values (40-502 mg C m⁻² d⁻¹) were low and not up to Arabian Sea values for the same season. Diatoms dominated the phytoplankton community and consisted of more number of genera compared to the Arabian Sea. Large colonies of the tunicate, *Pyrosoma*, which occurred at the surface as well as mid-depths could have consumed a part of the phytoplankton population. These results, albeit limited, have implications on the biogeochemistry of the region.

1. Introduction

The Arabian Sea (AS) and the Bay of Bengal (BOB) are twin seas caressing the western and eastern borders of the Indian subcontinent. The terrains look almost alike, both being enclosed in the north at similar latitudes. There are, however, many

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differences - the latter is partially connected to the Pacific through the Australasian seaways, contributing to physical, chemical and biological exchanges. Both lie in the tropics and are relatively small basins coming under the influence of the seasonally reversing monsoons. They form excellent crucibles to study the variability in the biogeochemistry of the oceans.

Differences between the physical and chemical regimes of AS and BOB arise mainly through (1) opposing trends in surface circulation during summer (southwest) and winter (northeast) monsoons (2) the BOB receives large quantities of fresh water from hinterland rivers (an imposing $1.6 \times 10^{12} \,\mathrm{m}^3 \,\mathrm{yr}^{-1}$ compared to $0.3 \times 10^{12} \,\mathrm{m}^3 \,\mathrm{yr}^{-1}$ in the AS, latter including the Indus; Subramanian, 1993) as well as oceanic precipitation making its upper layers less saline (annual mean <34 psu). The associated suspended sediment discharge into the Bay is estimated to be $1.4 \times 10^9 \,\mathrm{tonnes}$ compared to about $200 \times 10^6 \,\mathrm{tonnes}$ in the AS.

Under the aegis of the JGOFS (Joint Global Ocean Flux Studies) AS was targeted as one of the prime areas for investigations along with the Atlantic, Pacific and Southern Ocean. Thus, over the recent past we have acquired substantial amount of data sets from the AS (Burkill et al., 1993; Burkill, 1999; Baars, 1994; Lal, 1994; Krishnaswami and Nair, 1996; Gaur, 2000; Smith, 1998, 1999, 2000, 2001). On the other hand, after the International Indian Ocean Expedition (IIOE) during 1959-1965, despite many studies to understand the physical characteristics of the Bay of Bengal (for e.g., La Fond, 1957; Suryanarayana et. al., 1991; Murty et al., 1992; for more detailed reviews see Varkey et al., 1996; and Schott and McCreary, 2001), very few attempted to comprehend the water column chemistry (Sen Gupta et al., 1977; De Souza et al., 1981; Rao et al., 1994). Even fewer studies exist which address the biogeochemical characteristics of the Bay using collocated sampling and the overall information available remains scanty. For this reason, the Indian-JGOFS extended its studies from the AS to this region as a sequel, with an aim to cover seasonal and interannual changes and understand the processes which drive the biogeochemistry of the region (see the web site www.bobps.indian ocean.org/bobps).

BOB is traditionally considered to have poorer biological productivity compared to its western counterpart - the AS. Although the riverine flux may bring in nutrients, they are thought to be lost to the deep because of its narrow shelf (Qasim, 1977; Sen

Gupta et al., 1977; Radhakrishna et al., 1978). Comparatively heavier cloud cover during summer monsoon may be another reason for this. However, it is a cyclone-prone region and these episodic events are likely to churn-up the area, injecting nutrients to the shallow euphotic zone (shallow due to cloud cover and turbidity arising from sediment influx) and thereby enhance production in the upper layers.

In fact, it was postulated that higher surface production resulting from lower light intensities occurs in the BOB although the column production is much higher in the AS (Qasim, 1977). Primary productivity values ranging from 3.0 to 8.7 g C m⁻² d⁻¹ have been reported from the *inshore* waters of the east coast of India in June-July (Nair et al., 1973). Based on measurements during August-September 1978 chl *a* up to 50 mg m⁻² was reported inshore by Bhattathiri et al. (1980) and concluded that "the primary production of BOB is not less than that reported from the AS" (see also Pant,1992). Nonetheless, these values are exceptional, often referring to inshore blooms. On the contrary, most measurements are in effect less than 0.5 g m⁻² d⁻¹ along with very low chlorophyll *a* (usually <0.1 mg m⁻³ or <10 mg m⁻²).

The BOB Process Studies were aimed to understand (1) whether the riverine inputs influence the upper ocean physical and chemical characters and spatio-temporal variability in production (2) how the altered features of the BOB compare with the AS and its biogeochemistry. With these aims, we undertook the first BOB Process Study during summer monsoon from July 6 to August 2, 2001, covering two transects, one in the open sea (along 88°E) and another along the eastern coast of India (81 - 85°E; Fig. 1). It may be pointed out that the coastal track was in fairly deep waters (depth often exceeding 1000m) because of the narrow shelf.

2. Data and Methods

A Sea-Bird Electronics CTD (Conductivity-Temperature-Depth) was employed to obtain profiles at one-degree intervals and to 1000m depth on board the ORV *Sagar Kanya*. CTD salinity was calibrated against water samples collected simultaneously by a rosette sampler fitted with 30 litre Go Flo bottles and analyzed with a Guildline 8400 Autosal. The rosette was allowed a 1-minute stabilization before the bottles were closed to ensure that samples from desired depths were collected without any depth ambiguity. Surface meteorological parameters were measured continuously using the shipboard

automatic weather station. The sea surface temperature (SST) was measured using a bucket-thermometer by keeping it as close to the water surface as possible without breaking the water surface. Water samples from various depths were analyzed for nitrate and silicate with a SKALAR auto-analyzer.

Apart from the high resolution one-degree sampling for physical and chemical observations, five stations in the open ocean and four stations along the western margin (Fig. 1) were occupied, each for over a day, for measuring primary productivity and related parameters. Water samples collected from eight discrete depths (near surface, 10, 20 m and thereafter at 20 m intervals up to 120 m) by the CTD rosette sampler were used for measuring primary production (PP), chlorophyll *a* (Chl *a*) and for estimating phytoplankton cell counts from these nine stations. The JGOFS Protocols (UNESCO, 1994) were followed for measuring all parameters.

For PP measurements water samples drawn at pre-dawn from the above mentioned depths were collected into four 300 ml polycarbonate bottles (Nalgene, Germany) (three light, one dark) for each depth. One ampoule of NaH¹⁴CO₃ (Board of Radiation and Isotope Technology, Mumbai with a specific activity of 185 kBq) was added to each bottle and incubated *in-situ* using a mooring system, at respective depths for 12 hours from just before sunrise to half an hour after sunset. Incorporation of ¹⁴C was determined by filtering 100 ml of sample from each bottle through GF/F filters (25 mm dia, 0.7 µm pore size, Whatman, USA). The filters were transferred to scintillation vials and exposed overnight to HCl (0.5 N) fumes in a closed container. 5 ml of liquid scintillation cocktail (Sisco Research Laboratory, Mumbai) was added to each vial and the radioactivity was measured in a scintillation counter (Wallac 1409 DSA, Perkin Elmer, USA). Primary production rate was calculated and expressed as mg C m⁻³ d⁻¹, 12 h being considered as day (UNESCO, 1994).

Two replicates of 1.5 litre samples from the depths mentioned above were filtered through 47mm GF/F filters and deep frozen immediately at -20 °C. At the end of the cruise, all filters were transported to the shore laboratory in liquid nitrogen. Within a week, the chl *a* was extracted in 10 ml 90% acetone in the dark for 24 hrs in a refrigerator and its concentration determined fluorometrically (Turner Designs, USA).

For an assessment of generic composition and cell counts of phytoplankton (size >5µm), water sample from each depth up to 60m were fixed in Lugol's iodine (1%;

w/v) and 3% formaldehyde. Samples were stored in dark at low temperature until analyses. A settling and siphoning procedure was followed to concentrate samples from 250 ml to 20 ml. Two 1 ml replicates of concentrated samples were then examined microscopically in a Sedgewick-Rafter plankton counting chamber at 200x magnification. In addition to counting the phytoplankton, 1.5 to 3.0 ml subsamples from the samples collected for bacterial abundance measurements (formalin preserved) from similar depths were examined for autofluorescing cells retained on polycarbonate membranes (0.22 μm pore size). Autofluorescing picoplankters were enumerated using oil immersion objective on an epifluorescence microscope (Nikon E400, Japan) at 1000x.

3. Results

3.1 Physical features

Wind speeds, along 88°E, were on an average about 6 m s⁻¹ from 7°N to 13°N and increased further north to 10.5 m s⁻¹. Wind direction varied from west to northwest (Fig. 1). Atmospheric pressure showed a steady decrease towards the north from 1013 mb at 7°N to 1003 mb at 20°N. Except for the diurnal variability, air temperature did not show any significant variation, with an average value of 28.7°C. SST was on an average 28.4°C between 7°N and 9°N and increased to 29.5°C at 11°N. Mixed layer depth (MLD), calculated using the density criteria (Levitus, 1982), was in excess of 50 m at 12°N but shoaled under the influence of increasing freshening more or less gradually to less than 4m at 20°N (Fig.2, and also shaded region in Fig. 6).

The vertical thermal structure in the upper 300 m showed oscillation within the thermocline (Fig. 2a). An upheaval of isotherms was noticed between 7°N and 11°N while north of it the isotherms showed a gradual shoaling towards 20°N. In the former case, the 28°C isotherm shoaled from 80m to 20m and thus the upper thermocline showed a vertical displacement of about 60m, which tapered off towards the lower thermocline. This was clearly a signature of a subsurface cold core eddy, seen below 15m depth, which depressed the ambient temperature at 60m (27°C) by about 5°C. Below 300m, the thermal structure did not show any special features.

The vertical salinity structure showed strong gradient in the upper 100m depth (Fig. 2b). In the surface layer, salinity was about 33.5 psu (practical salinity scale) between

7°N and 15°N but reduced rapidly towards north reaching a low of 27.6 psu at 17°N. The salinity gradient in the upper 50m at 7°N was about 1.5 psu while that at 20°N was about 7 psu, indicating the strong signal of freshwater influx in this region. Below 100m depth, the vertical salinity distribution showed almost homogeneous waters down to 1000m (data not shown) with a salinity of 35 psu.

Along the coast, wind speeds were on an average 4 m s⁻¹ except at 12°N and 18°N where it was about 9m s⁻¹. Wind direction varied from south to north-northwest with average winds being in a southwesterly direction (Fig. 1). As was the case of open ocean section, the atmospheric pressure also showed a decrease towards the north. Air temperature did not show any significant trend except for diurnal variability, with an average value of 28.5°C. SST exhibited a 1°C drop from 11°N to 20°N. Coastal SSTs were on an average 28.9°C and were higher than those at the open ocean stations of the corresponding latitude. MLD, which was 25m at 11°N shoaled northwards to almost imperceptible levels (Fig.3, and also shaded region in Fig.6).

The thermal structure in the upper 300m along the coastal section also exhibited oscillations within the thermocline (Fig. 3a) as was the case with the open ocean transect. A noteworthy feature was, again, an uplifting of isotherms centered at 17°N, 83.5°E. The 28°C isotherm shoaled from 60m to 10m (Fig. 3a). This vertical displacement of about 50m in the upper thermocline depressed the ambient temperature by about 5°C. Although we did not observe any upwelling signature in this section in terms of cold and high salinity waters in the upper layers, this could be because our western boundary stations sampled waters much farther away from the coast/shelf.

Vertical salinity structure in the upper 300m along the coast also showed strong salinity gradient in the top 100m depth (Fig. 3b) and homogeneous waters with 35 psu salinity below it and up to 1000m. But unlike the open ocean, the salinity gradient south of 15°N was weaker (0.5 psu as compared to 1.5 for the open ocean). In the surface layer the average salinity was 33.7 psu up to 16°N but reduced northward, with the lowest value of 29.6 psu at 19°N.

3.2 Nutrients

Vertical distribution of nitrate showed that in general the upper 30m was devoid of nitrate and this situation deepened up to 50m between 12°N and 15°N (Fig. 4a). The

 $1\mu M$ isopleth showed shoaling under the influence of the subsurface cold core eddy and towards the north the shoaling was concurrent with that of the isotherms (Fig. 2a). In general, nitracline was situated between 50 and 100m depth. A second region of strong gradient (of about 30m thickness) was seen around 200m. Silicate distributions were also similar to that of nitrate, except for a high concentration of more than 2 μM in the north (Fig. 4b). This indicated the signature of fresh water influx from the north.

Nitrate distribution along the coastal section showed that the upper 40 m was devoid of nitrate except in the north (Fig. 5a). However, under the influence of the cold core eddy signature seen in the thermal structure (Fig. 3a), we noticed shoaling of nitracline as was evident from the 1 μ M isopleth which shoaled up to 30m between 16°N to 18°N. Below the nitracline, the pattern remained the same as that of the open ocean. Silicate distribution was also similar to that of nitrate, except for the presence of high surface silicate (> 4 μ M) in the north (Fig. 5b) as in the case of open ocean section. Phosphates were generally at undetectable levels in the upper 30m throughout the area of study.

3.3. Biological characteristics

Concurrent with nutrient distribution, surface chl *a* was poor (0.06 to 0.28 mg m⁻³ in open waters and 0.06 to 0.16 mg m⁻³ towards the coast, Fig. 6a,b). Satellite (Sea-Wifs) derived pigment concentrations for the monthly composition of July 2001, (T. Suresh, pers. comm.) also showed similar surface values of ranging from 0.1 to 0.3 mg m⁻³ in the sampled region. Higher surface values were towards north and so were the column integrated values (range 10-20 mg m⁻², Fig. 7). A subsurface chlorophyll maximum (SCM) was encountered at most stations (Fig. 6 a,b) usually below the mixed layer and associated with the nutricline (~40m). On an average, about 37% of chl *a* occurred in the MLD while 28% occurred at the SCM depth. Only in the northern most open ocean station (20°N) a decrease in chl *a* with depth in the upper layers was observed where it was uniformly 0.28 mg m⁻³ upto 10m depth. It might be that the low salinity associated with a virtually non-existent mixed layer caused this. There were no clear differences between open and coastal waters, except that 20% of chl *a* occurred below 80m compared to 13% in open waters.

Column integrated PP was between 90 and 220 mg C m⁻² d⁻¹ along 88°E, but relatively higher range was obtained along the coastal transect (328-520 mg C m⁻² d⁻¹) except for a low 40 mg C m⁻² d⁻¹ at 15°N (Fig. 7). PP within the mixed layer was about 54% of the total while below 80 m it was insignificant (8% in both coastal and open waters). About 26% of the PP was associated with the SCM. The productivity to chl a ratio was between 10 and 20 although a few higher values (up to 40) occurred at some coastal stations (Fig. 7). A significant correlation (r = 0.58; P < 0.001; n = 72) between chl a and primary productivity rate was observed (Fig. 8) showing that most of the productivity was related to the active chl a present in the water column.

Along the open ocean the phytoplankton cell counts were the highest at 20° N ranging from $13.8 \times 10^{3} \, \Gamma^{1}$ at surface (corresponding to chl a estimates) to $0.6 \times 10^{3} \, \Gamma^{1}$ at 60m. They showed a general decrease southward, with their lowest abundance at 15° 88°E (Fig. 9). At stations south of 15° N, their counts were higher in the upper 10m and decreased drastically below this. Along the western margin the highest counts were observed from samples collected at 19° N 85° E and the lowest at 15° N 81° E (Fig. 9). Their counts kept increasing with increasing depth at 12° N 81° E for reasons unknown. Similar to discrete depth variations, the column integrated numbers clearly indicated their preponderance in the north. In general, the phytoplankton counts, chl a concentration and primary productivity ratio along both transects showed co-variation. Picoplankton abundance was generally higher along 88° E with a range of $0.24 \times 10^{7} \, \Gamma^{1}$ to $18.5 \times 10^{7} \, \Gamma^{1}$ compared to coastal areas except a high $(9.4 \times 10^{7} \, \Gamma^{1})$ at 40m) in the north at 19° N 85° E (data not shown).

Diatoms dominated the phytoplankton population both in open and coastal waters (>90%, Table 1, depicted from 2 representative stations). *Biddulphia, Chaetoceros, Thallasiothrix, Thallassionema, Schroderella, Navicula* and *Nitzchia* were the common genera. A few genera of dinoflagellates, usually *Peridinium* and *Ceratium* occurred.

Zooplankton biomass (collected at mid-day and midnight from different strata up to 1000m using a Hydro-Bios Multiple Plankton Net with a mesh size of 200 µm) at the 9 PP stations was generally concentrated in the MLD and rapidly diminished with depth especially below the lower thermocline (82% of the biomass in oceanic waters and 59% in coastal areas was above the thermocline; for sampling strategy and biomass conversion to carbon see Madhupratap et al., 2001). Day-night biomass did not show

any particular pattern. The biomass in the upper 1000m varied usually from 18 to 400 m mol C m⁻² in open waters and 12 to 1070 m mol C m⁻² in the western boundary. But, we could not realistically estimate the biomass at most of the open ocean station and one along the coast (11°N, 81°E) because huge colonies of the tunicate *Pyrosoma* occurred which could not be separated from other zooplankton. These large colonies also occurred without any day-night pattern (given that the ship drift over 12 hr would have sampled different, but adjacent areas). Pyrosome colonies were seen at times in the surface and inexplicably in layers below up to 500m depth and the biomass at these depths (> 1000 m mol C m⁻²) led to exceeding values in comparison to conventional estimates. Otherwise, the zooplankton were dominated by copepods – *Paracalanus, Clausocalanus* and *Eucalanus* (calanoids) being the common genera plus the carnivorous *Oithona* (Cyclopoid) and *Oncaea (Poecilostomatoid*) comprising the composition.

4. Discussion

The investigation was conducted during the summer monsoon when the rivers of India are in spate and discharge large quantities of freshwater into the adjacent seas, particularly the Bay of Bengal. The rainfall during the year 2001 showed a normal summer monsoon season (Indian Meteorological Department; www.tropmet.res.in). The effects of the river plume were evident in the northern regions where the highest discharge arises out of the Ganges - Brahmaputra - Irrawadi rivers (Subramanian, 1993), reducing the surface salinity to ca. 28 psu or lower.

A significant feature observed from the thermal structure, both along the open ocean and western boundary was the presence of thermocline oscillations and cold core eddy signatures. Oscillations similar to present observations were recorded by Rao et al. (1983) from a time series measurement at 7.5°N and 87.5°E during May 1979. The formation of eddies was also noticed close to the western boundary in July 1984 (Babu et al., 1991). Apart from these, we also observed a northward shoaling of isotherms north of 12°N along the open ocean section. Under the influence of these eddies not only the subsurface cold waters but nutrients as well were pumped into the upper subsurface layers. Consistent with the northward shoaling of isotherms in the open ocean section, nitrate as well as silicate isopleths also showed shoaling. However, the

lense of 2-4 µM silicate in the surface layers, detached from the subsurface, in the northern most region sampled in both sections showed that this was a feature much closer to the coast in the north and did not arise out of upward pumping. In fact, the effects would be more in the inshore/shelf waters which we did not sample. Results obtained by Rao et al., 1994 in March-April (a dry season) and December (cyclone-prone) from shelf to offshore did not show presence of nitrate in the upper layers and concluded that rivers flowing into the Bay might not contribute much to the inorganic nutrient pool. This is in confirmation with earlier results obtained by De Sousa et al. (1981) and Rajendran et al. (1980) during south west monsoon and that a considerable part of the terrigenous humus is lost at its confluence with the sea (Sen Gupta et al., 1977). The upper low salinity layer caused distinct stratification in summer resulting in generally shallow MLDs, despite favorable wind speeds which should have resulted in deeper mixed layers.

Thus, both eddies and shoaling isotherms were capped by the low saline layer leading to a failure of their surfacing and prevented nutrients reaching the upper (40m or so) layers. Neither did the river plume bring in appreciable nutrients southward or these were consumed within the estuary or inshore waters. Localised, mild upwelling along the southern part of the western boundary during summer had been reported by Murthy and Varadachari (1968) and De Sousa et al. (1981). Later Shetye et al. (1991) suggested upwelling along the east coast of India during summer, in an approximately 40 km wide band, but they also showed that river discharge in the north overwhelms signatures of upwelling.

The biological features were consistent with the observed physical and chemical regimes. The low chl *a* and PP dovetail with low ambient nutrients. Reduced irradiance and heavy sediment discharge in this season shallows the euphotic depths to 6m (inshore) and to about 20-40m (open ocean) in this season (Qasim, 1979). Even cyclone created perturbations are bound to result in the same. We are yet to measure these from other seasons, nevertheless, our first conclusions are that the BOB is less productive in

early summer in comparison to the AS.

Earlier reports of Qasim (1979), Bhattathiri et al. (1980) and Devassy et al. (1983) indicate that chlorophyll a concentrations varied widely without showing any geographic or seasonal patterns. Radhakrishna et al. (1978) reported surface chlorophyll a to range from 0.084 to 1.67 mg m⁻³ and euphotic column values from 8.63 to 28.45 mg m⁻² during 1976 summer monsoon. For the 1978 summer monsoon, the column chlorophyll a in euphotic zone ranged from 1.28 to 33.72 mg m⁻² in the offshore waters (Bhattathiri et al., 1980) and from 0.01 to 1.01 mg m⁻³ in the surface waters. Concentrations were in the range of 2.11 to 23.60 mg m⁻² during 1977 monsoon months and the surface concentrations varied from as low as 0.03 to a high of 1.04 mg m⁻³ (Devassy et al., 1983). The high value observed by them was attributed to a diatom bloom predominated by *Thallassiosira subtilis*. The observed range of surface chl a in the present study was low compared to some of the earlier reports and was in the range of 0.06 to 0.28 mg m⁻³. The SCM seen at most stations is consistent with observations from the Bay during May-June, 1996 (Murty et al., 2000); the pigment concentrations at the SCM were found to be an order of magnitude higher than those at the surface. The range observed was similar to ours being 0.01 to 0.2 mg m⁻³. However, Gomes et al. (2000) report that Chl a and production rates could go as high as 53 mg m⁻² and 4.5 g C m⁻² d⁻¹ in March-April, a season with very less run-off. But, Sen Gupta et al. (1977) show that there are no detectable nitrates in the upper layers of the BOB along the western region in this season as well. During summer monsoon the values of chl a were higher up to 92 mg m⁻² but PP on the other hand remained low (0.3 g C m⁻² d⁻¹). Gomes et al. (2000) attributed the reason for this to the cloud cover.

Historical measurements of PP from the Bay date back to *Galathea* Expedition of the 1951 followed by the *Anton Brunn* expedition. These measurements show values of 0.1 to 0.3 mg C m⁻² d⁻¹ in the deep and 0.01 to 2.16 g C m⁻² d⁻¹ in shelf region. Later values reported from offshore regions are from 129.99 - 329.45 mg C m⁻² d⁻¹ (av. 219.79) during August-September 1976 (Radhakrishnan et al., 1978) and from 180 to 2200 mg C m⁻² d⁻¹(av. 740 mg C m⁻² d⁻¹) during Aug-Sept 1978 (Bhattathiri et al., 1980). In the slope region along the western margin of the Bay, the PP during August-September, 1976 was in the range of 49.66 – 606.37 mg C m⁻² d⁻¹ (av. 315.43 mg C m⁻²

d⁻¹) and during Aug-Sept 1978, it was 120 to 3410 mg C m⁻²d⁻¹ (av. 1280 mg C m⁻² d⁻¹). However, these were made before trace-metal clean techniques were available and a recalculation shows that the PP to chl *a* ratios are mostly lopsided often exceeding 250 and up to >2500 (except Radhakrishna et al., 1978). Thus we do not reconcile totally with available earlier data, but concede that inshore/coastal measurements are lacking in our observations.

With clean-techniques and standardization of *in-situ* incubation protocols, reliable measurements of PP during the last decade from various parts of the world oceans are available (see Letelier et al., 1996, Barber et al., 2001). Since the present data set gives the first *in-situ* measurements from the Bay, more recent southwest monsoon data available from the Arabian Sea are compared in Table 2. Clearly the PP in the Bay is much lower than those observed in the Arabian Sea during this season. The PP in the Bay during southwest monsoon of 2001 was more comparable to those observed during the oligotrophic Spring-Intermonsoon in the Arabian Sea when SSTs are high (28-29°C and comparable to the present study) and mixed layer shoals and attains more or less the characteristics of a Typical Tropical Structure (Madhupratap et al., 1996, 2001). Unlike the BOB, in the AS strong upwelling along the western boundary and in the open ocean during southwest monsoon (Prasanna Kumar et al., 2001) bring nutrients up into the euphotic zone enhancing productivity.

The numerical abundance of phytoplankton ranged from 0.41 x 10⁸ to 7.1 x 10⁸ m⁻² during the present study. Somewhat paradoxically, this range is comparable to those reported earlier from the central and eastern Arabian Sea (0.3 x 10⁸ to 10 x 10⁸ m⁻²; Sawant and Madhupratap, 1996) during southwest monsoon of 1995. Similarly, contribution of diatoms to the total phytoplankton was significantly large in both areas. Nitrate rather than silicate seem to control PP in both regions. However, the number of genera of diatoms found in BOB (26) was higher than those found in the AS (9) thus showing higher diversity. There could be a reason for this; as food becomes scarcer, as may be the case in BOB, allopatric or even sympatric speciation is found in deep-sea copepods leading to higher diversity (Madhupratap et al., 1990). This plus salinity variations in the Bay might result in niche separation and we already know that composition of species of fish in this area is quite different from that of AS (CMFRI, 1995). It is speculative, but at the moment we do not have better explanations. The

population of autotrophic dinoflagellates was low in the Bay. They formed $\sim 5\%$ of the total phytoplankton community in the oceanic waters (Table 1). Higher primary productivity value associated with high chlorophyll a concentration at northern most coastal station (19°N 85°E) was reflected in the total phytoplankton cell counts as well.

Zooplankton biomass obtained from the upper 1000m of the AS (Madhupratap et al., 2001) during southwest monsoon (94 m mole C m⁻²) is not strictly comparable with those from present study because of the large pyrosome colonies. Otherwise the common genera of copepods present in the two regions were more or less same. Incidentally, the pyrosomes are voracious mucous/filter feeders and are a counter part of the salp swarms very frequently found in the AS during summer and winter. This could also have led to the observed reduction in phytoplankton biomass through grazing.

All these bring us to some other questions pertaining to the biogeochemistry of the BOB. The fact remains that the productivity of the BOB remains an enigma with conflicting reports over seasons and areas. Obviously, this single report covering a very short period is not resolving it. But, it would seem that it is consistent with observed earlier reports on nutrient data. Some upwelling could happen inshore in some areas as indicated earlier and lead to higher productivity. But the overall annual picture is yet to emerge.

An oxygen minimum zone (OMZ) between 100 and 500m, less thick compared to the AS occurs in the BOB (Olson et al., 1993). One of the reasons attributed to cause the OMZ in the AS is the flux and decomposition of organic particles brought into the deep derived from the overlying productive waters. If the production in the BOB is lower, how does this happen in these waters? And, BOB does not make significant contribution to the water column denitrification which arises as a result of OMZ unlike the AS (see Rao et al., 1994, for a discussion). Another puzzle is that the average annual fluxes into the deep are comparable both in AS and BOB. However, lithogenic fluxes are more in the Bay and increase in fluxes are associated with increase in river discharge (Ittekkot et al., 1991; Ramaswamy and Nair, 1994). It may be that biogenic matter is more rapidly scavenged along with those of terrigeneous origin and ballast the material resulting in faster sedimentation to the deep (Ittekkot et al., 1992; Kumar et al., 1998). And thus, it may be fluvial matter (organic and undecomposed matter as

indicated by Sen Gupta et al., 1977) which might be contributing more to the fluxes. Some of these questions along with interannual variability will be addressed in more detail in the forthcoming cruises of BOB Process Studies.

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Legends to figures

- Fig. 1. Location map showing the stations along open ocean (88°E) and coastal track. Dark circle indicate 1-degree CTD stations while hollow circle are the biological stations for *in-situ* incubation for primary production. The arrows are the observed wind vectors at stations.
- Fig. 2. Vertical section of (a) temperature (°C) and (b) salinity (psu) along open ocean (88°E) track.
- Fig. 3. Same as Fig.2 but along coastal track.
- Fig. 4 Vertical section of (a) nitrate (μm) and (b) silicate (μm) along open ocean (88°E) track. Note the change in contour interval of dashed line.
- Fig. 5. Same as Fig. 4 but along coastal track.
- Fig. 6.Vertical profiles of primary production (solid circles) and chlorophyll *a* (open circles) in (a) oceanic (b) coastal stations (shown by lat., long.). Mixed layer depths are shown as shaded areas.
- Fig. 7. Integrated concentrations of chlorophyll *a* (chl *a*, mg m⁻²), primary production (PP, mg C m⁻² d⁻¹) and PP to chl *a* ratios over 120m in the oceanic and coastal waters.
- Fig. 8. Relationship between chlorophyll a (mg m⁻³) concentrations and primary production (mg C m⁻³ d⁻¹).
- Fig. 9. Depth distribution of phytoplankton cell density (nos. x 10³ l⁻¹) in the Bay of Bengal during July-August 2001.

Table 1 Generic composition of diatoms and dinoflagellates at two representative stations (0-60m; 17°N 88°E and 19°N 85°E). Numbers are percentages derived from the average counts from 0-60m.

Phytoplankton generic	1701.005 1001.055	
composition	17N 88E	19N 85E
Total cell counts (l ⁻¹)	4594	10294
Diatoms	<u> </u>	
Amphora	0.22	1.53
Asterionella		3.96
Bacteriastrum	0.17	1.55
Biddulphia	3.78	14.67
Chaetoceros	28.30	
Climacosphenia	1.10	
Corethron	0.23	2.32
Coscinodiscus	4.69	0.45
Ditylum	0.88	1.27
Eucampia	0.36	
Fragilaria	1.14	
Grammatophora	0.35	0.07
Gyrosigma		1.53
Leptocylindrus	0.53	
Melosira	2.33	1.52
Navicula	6.51	16.35
Nitzchia	5.65	1.53
Psedonitzchia		8.02
Rhizosolenia	7.90	
Schroderella	0.07	8.06
Skeletonema		0.14
Surirella	0.17	0.58
Thallassionema	4.26	30.57
Thallassiothrix	23.82	2.36
Thalassiosira	0.30	1.24
Unidentified	1.37	
Dinoflagellates		
Ceratium	1.25	
Dinophysis		0.21
Goniaulax	0.44	
Peridinium	3.53	2.01
Prorocentrum	0.66	0.07

Table 2.

Primary productivity and chlorophyll *a* values from the Arabian Sea and Bay of Bengal determined during southwest monsoon by following JGOFS protocols.

	PP (mgCm ⁻² d ⁻¹)	Chlorophyll <i>a</i> (mg m ⁻²)	Reference	
Arabian Sea				
Shelf	440-1760	16-88	Bhattathiri et al. (1996)	
	800-2800	33-54	Veldhuis et al. (1997)	
	449-2668 [§]	0.63-1.66 mg m ⁻³ *§	Owens et al. (1993)	
Offshore	770	34-44	Bhattathiri et al. (1996)	
	770-1782	26-60	Prasanna Kumar et al. (2001)	
	1320-1590	0.09- 1.31mg m ⁻³ *	Barber et al. (2001)	
	236-533 [§]	0.17-0.4 mg m ⁻³ *§	Owens et al. (1993)	
Bay of Bengal				
Shelf	39.7-502.0	11-19	This study	
Offshore	89.4 -220.7	9-11	This study	

^{*}Column values not available

[§] Values for September-October

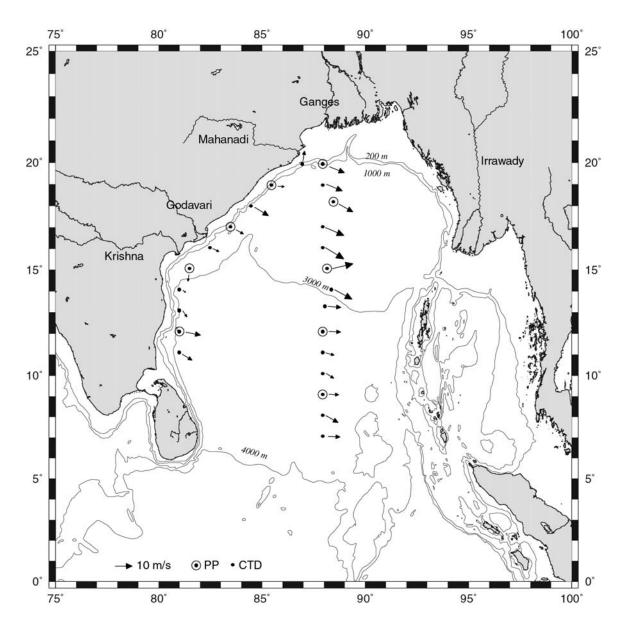


Fig.1

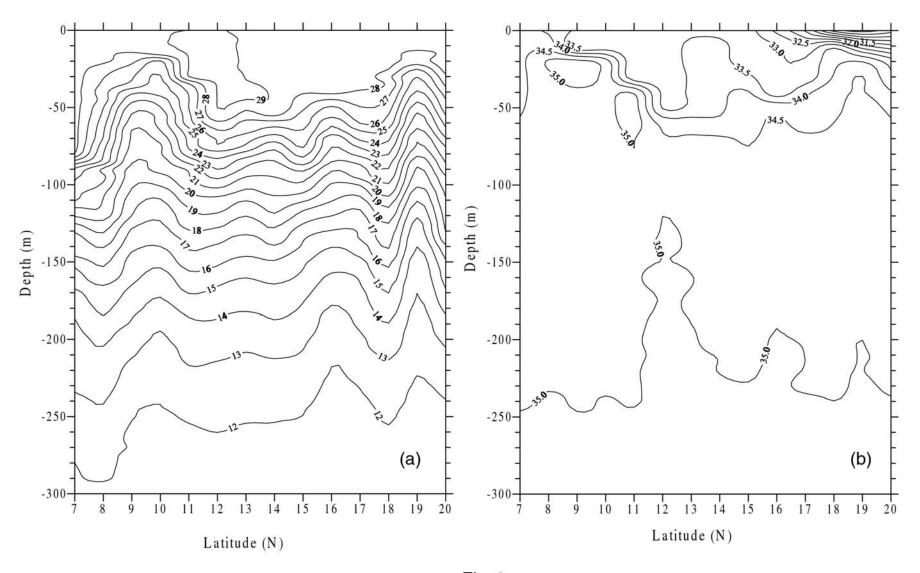
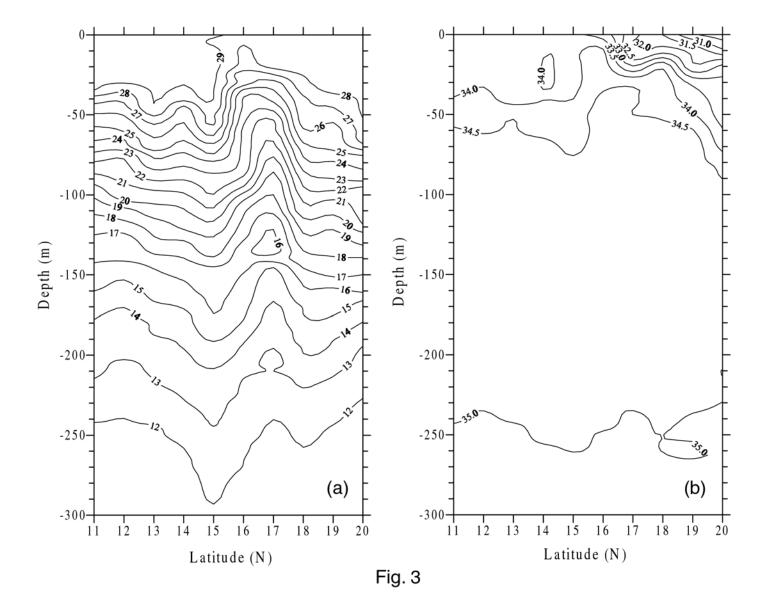
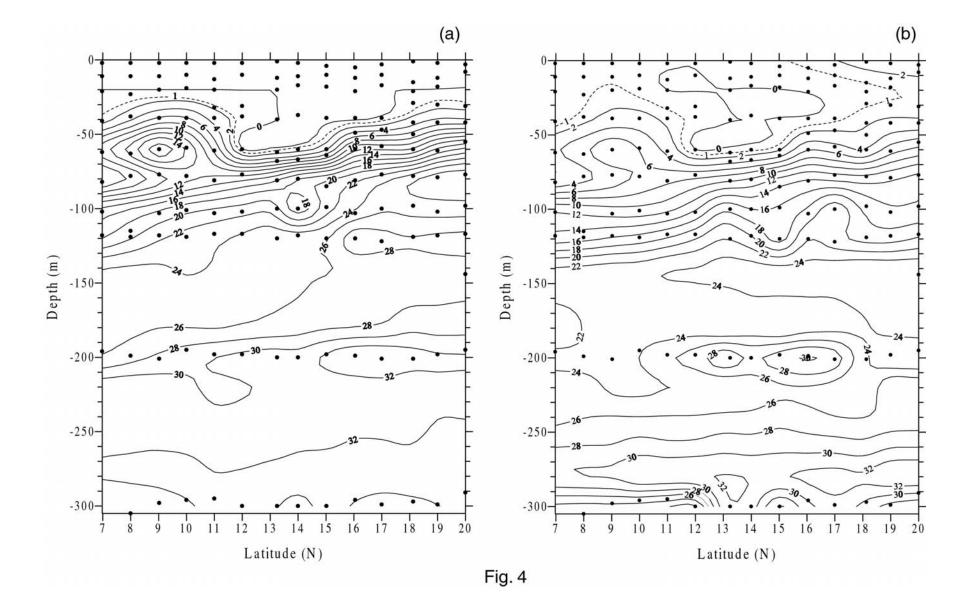
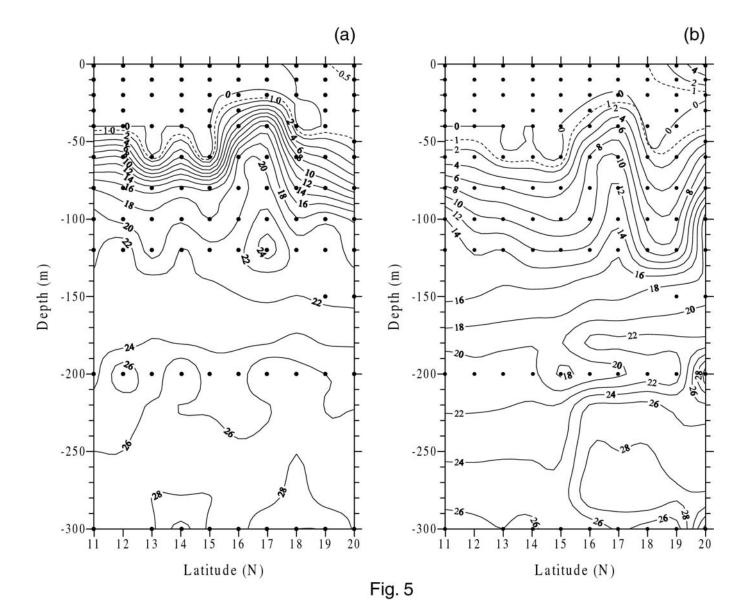
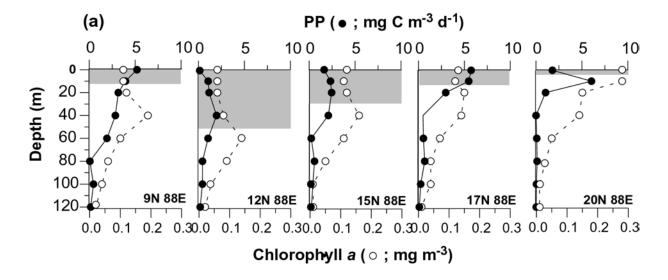


Fig. 2









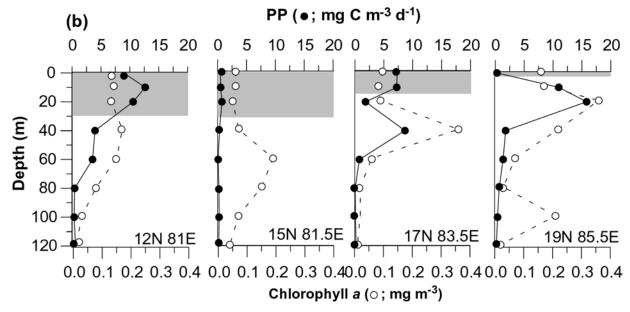


Fig.6

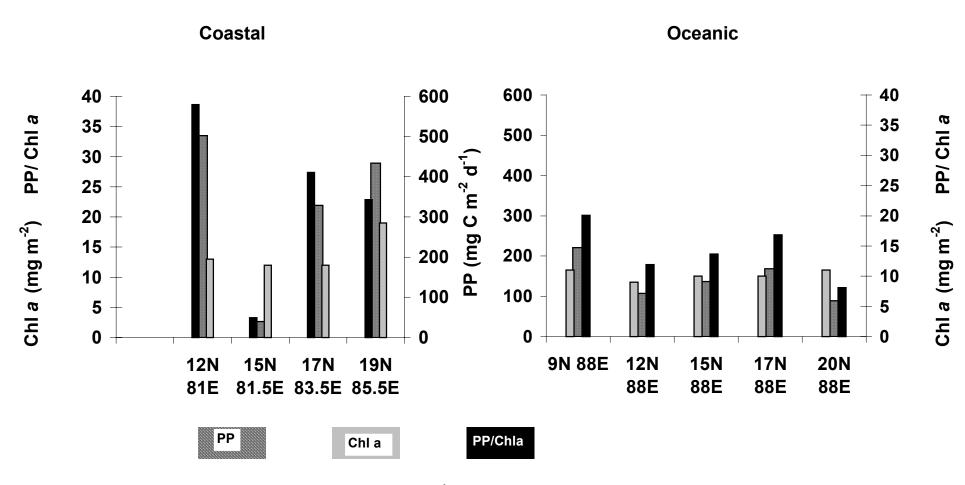


Fig.7

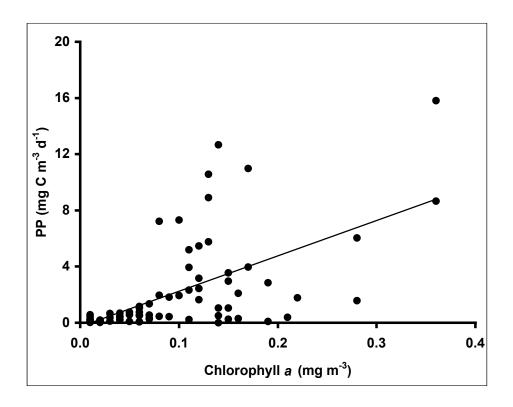


Fig. 8

