This study has helped in understanding how the efficiency of biting and feeding is increased in the landleeches. Firstly, the distribution of denticles is such that the smallest denticles are situated on the outer peripheral end of a jaw, where microincision on the host-. surface is initiated (figure 2). When a jaw is moved in a centrifugal manner, this small wound is slowly deepened with the help of larger denticles, found towards the central oral aperture; and widened with the help of the wedge-shaped jaws bearing a broad base and attenuated edge. Secondly, a typical denticle in the land-leech is considerably curved outwards, a fact seen neither in the Indian cattle-leech, P. granulosa, nor in the European leech, H. medicinalis. Lastly, the denticles are distinctly sharper and more acute-tipped than those of P. granulosa. The presence of such a denticular curvature and acute tip obviously adds to the sawing efficiency of the jaws of the land-leeches during microincision.

The present study, supported by light microscopic work, has shown that a large number of salivary ductules pass through a denticle and open at its tip. In addition to such an intra-denticular salivary discharge, a few openings can also be seen around the denticles. In short, a denticle not only pours saliva but is bathed by it also during microincision. A similar intra-denticular salivary discharge has been reported by Damas² in *H. medicinalis.*

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- 1. Damas, D, Arch. Zool. Exp. Gen., 1962,101, 41.
- 2. Damas, D., Arch. Zool. Exp. Gen., 1974,115, 279.
- Mann, K. H., Leeches (Hirudinea): Their structure, physiology, ecology and embryology, Pergamon Press, Oxford, 1962, 201.
- Mill, P. J., *Physiology of annelids*, Academic Press, London, 1978, 683.
- Yanagisawa, H. and Yokoi, E., Proc. Imp. Acad. Jpn, 1938, 14, 69.
- Bhatia, M. L., *Hirudinaria*, Emkay Publications, Delhi, 1977, 150.
- Dev, B., Maitra, S. C. and Shipstone, A. C, 13th Ann. Conf. Elec. Micr. Soc. India, Bangalore, 1981, B.6/B-105.
- Harding, W. A. and Moore, J. P. The Fauna of British India including Ceylon and Burma, Hirudinea, Taylor and Francis, Red Lion Court, Fleet Street, London, 1927, 302.

- 9. Stammers, F. M. G., Parasitology, 1950, 40, 237.
- Worth, B.C., J. Bombay Nat. Hist. Soc., 1951, 50, 423.

CHARACTERIZATION OF PURPLE AND GREEN PHOTOSYNTHETIC BACTERIA ISOLATED FROM THE LAGOON OF AGATTI ATOLL (LAKSHADWEEP SEA)

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PHOTOSYNTHETIC bacteria comprise green and purple bacteria which are different physiologically but with common anoxygenic photosynthesis using only one photosystem. They are different from cyanobacteria in that the latter group will carry out oxygenic photosynthesis using two photosystems¹. The contribution of photosynthetic bacteria to primary production in some environments can be very high and it varies from 20 to 85 % of the total daily production in some lakes. These bacteria are important not only as food for secondary production but also in the removal of toxic sulphide through anaerobic photooxidation in the process of CO₂ assimilation². Not much information is available on these bacteria in tropical waters except one report on the photosynthetic non-sulphur bacteria of Rhodospirillaceae in India³. The present communication is the first of its kind to report the occurrence and characterization of photosynthetic sulphur bacteria in the tropical waters.

During the 147th cruise of R.V. Gaveshani (5-25, January, 1985) routine microbiological parameters were studied in some of the Lakshadweep islands. While working in Agatti Island (Lat. 11°50'N and Long. 72°11'E) extensive areas of pink coloured sediments were noticed in the lagoon. These were found with decaying beds of sea grass. Since red water phenomenon in marine environment due to bacteria has been studied, and described in many coastal lagoons and lakes⁴⁻⁷, an attempt was made to isolate photosynthetic bacteria from the pink sediments and characterize them. Sediment samples were collected aseptically and about one gram of sample was added to screw capped culture tubes (25 ml cap) containing Pfenning's medium⁸. The tubes were incubated at room temperature (28 \pm 2°C) for 7 days under natural

diffused su n ligh t On e se t o f tube s wa s als o sup plemente d wit h 0.5 g of sterile boile d egg per tube t o test for photoorganotrophi c Rhodospirillaceae. At the end o f the incubatio n period, som e tube s develope d intense purpl e viole t colouratio n an d som e other s green. In addition, some tube s develope d bot h purpl e and green colouration. Bacteri a from these tube s were subculture d an d subsequently purified b y seria l dil ution o n shak e agar.

It was observe d that t identical type so f purple bacteria develope d bot h o n organi c an d inorgani c media. The development was, however, faster in the organic medium . Th e cells tende d t o grow i n aggre gates on the walls of the tubes . Microscopi c examin ation of purple prototrophi c bacteri a from inorgani c medium revealed that the cells were elongate (4-6 μ m x 2-3 µm), motile, wit h sulphu r granul e inclusions. When the sulphid e content of the culture medium was in excess, cell s sometime s gre w upt o 8-1 0 μ m \times 3-4 μ m size. Organi c mediu m als o yielde d simila r cells. Absorptio n spectru m o f whol e liv e cell s sus pended i n 143 % sucros e solution ⁹ take n i n a BECKMAN-D U 6 scannin g spectrophotomete r showed that there were three peaks i.e. 805 run, 830nm and 863 nm i n th e infrare d regio n and two peak s i.e. 450 n m an d 59 0 n m i n th e visibl e region . Th e methanolic extract s of cells showed peak s at 365, 608 and 772 nm.

Green phototrophi c bacteri a consiste d o f non motile spherica l cells measurin g $1-1.5 \,\mu$ m in diameter. Sometimes the cells were found in chains. The absorp tion spectru m of whole live cells suspende d in sucros e solution showe d marke d peak s at 749, 486, 423 an d 380 nm. The methanoli c extracts of cells showed peaks at 666 and 435 nm.

When the character s of these culture s were compared with those of photosynthetic bacteria described by Trupe r and Pfennig⁹, the purple bacteria could be identified a s *Chromatium* spp. The spectral characteristics of the whole cells and their methanolic extracts prove that these strains contain bacteriochlorophyll'a'. The peak s observe d a t 450 nm and 590 nm in the spectrum of whole cell s could be attribute d t othe carotenoid pigments, most probably rhodopinal. The purple viole t colou r of cell suspension n together with the spectral characters, cell size and ability for photo-autotroph y permit the m to be tentatively identified as *Chromatium violascens*.

The possibilit y o f identifying th e present strai n a s either *Chr. warmingii* o r *Chr. buderi* which are als o purple-violet in colour, i s rule d out, becaus e thes e species hav e large r cel l siz e an d becaus e o f thei r inability to utilize compound s othe r tha n acetat e and pyruvate.

The phototrophi c green bacteri a could be identified as Prosthecochloris spp. The spectral characteristic s of live cell suspensions and methanolic extracts show that they contai n predominantly bacteriochlorophyll'c'. The peaks at 486 and 423 nm could be attribute d to the predominan t acarotenoid, chlorobactene⁹. The strain isolated in the present study is different from the marin e Prosthecochloris sp . describe d b y Matheron which exhibite d maximu m absorption at 460 nm and 750 nm in in vivo suspension and at 435 nm and 673 nm in methanoli c extracts. The identit y of the strain as Prosthecochloris is furthe r supporte d b y th e fact that they formed consorti a with sulphat e reducing bacteria in the primary isolation tubes, which is a characteristic feature o f thi s bacterium . Wit h th e availabl e data , this strain coul db e identifie da sa varian to f Prosthecochloris aestuarii which is the only green coloure d specie s s o fa r reported ⁹.

Experiments t o estimat e the contribution of these purple and green bacteria to primary production under photoorganotrophic and photoautotrophic conditions are under way.

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- 1. Avron, M., Curr. Top. Bioenerg, 1967, 2, 1.
- Pfennig , N., In: *The photosynthetic bacteria*, (eds) R. K. Clayto n and W. R. Sistrom, Plenum Press, New York, 1978, 3.
- 3. Karanth, N. G. K., Nair, S. and Lok a Bharathi, P. A., *Indian J. Mar. Sci.*, 1977, 6, 94.
- 4. Taga, N, Inf. Bull. Planktol., (Jpn), 1967, 219.
- 5. Truper , H. G. and Genovese , S., *Limnol Oceanogr.* 1968, 13, 225.
- Matheron , R., In: Contribution a 'I'etude ecologique, systematique et physiologique des Chromatiaceae et des Chlorobiaceae isolees de sediments marins. Thesis (Marseilles) 1979.
- Cohen, Y., Krumbein, W. E. and Shilo, M. *Limnol* Oceanogr, 1977, 22, 609.
- 8. Pfennig, N, I Abt Orig., 1965 1, 179 and 503.
- Truper, H. G. and Pfennig, N, In: *The prokaryotes*, (eds) M. P. Starr, H. Stolp, H. G. Truper, A. Balous and H. G. Schlege I (Springer-Verlag, Ne w York) 1981,299.