

Antibacterial Activity of Marine *Bacillus* Substances against *Vibrio cholerae* and *Staphylococcus aureus* and *In Vivo* Evaluation Using Embryonic Zebrafish Test System

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Cell free extracts of *Bacillus subtilis* subsp. *spizizenii* and *B. thuringiensis* isolated from hydrothermal vent region of Azorean Island Faial, North Atlantic Ocean showed antagonistic against *Vibrio cholera* serogroup 01 and *Staphylococcus aureus*, respectively. The *in vitro* inhibition of infectious pathogens by both *B. subtilis* subsp. *spizizenii* and *B. thuringiensis* was concentration dependent. The result of sensitivity to protease test showed considerable reduction of antibacterial activity for both *Bacillus* strains, indicating the proteinaceous nature of the antibacterial substances. Further, our results on antibacterial substance of heat resistant, proteinaceous nature and bacteriostatic mode of action was evaluated for the toxicity studies using zebrafish animal model, an alternative to the use of antibiotics test against infectious diseases. *Bacillus* antibacterial extracts showed no toxic effects with the *Staphylococcus aureus*-zebra fish embryo infection studies.

Key words: Antagonistic, bacterial disease, marine bacteria, Pathogen, zebrafish

The production of antimicrobial substances by marine bacteria isolated from different locations in the sea has been known for long time^[1,2]. But the usage of marine bacteria obtained from hydrothermal vent regions as a potential antibacterial agent against pathogenic disease was not much studied. In our earlier study we had reported several bacterial strains isolated from hydrothermal vent regions of Dom João de Castro Seamount (DJCS) located in the North Atlantic ocean, between the Azorean Islands of São Miguel and Terceira (38°13.3'N, 26°36.2'W), lies on the Terceira Rift^[3] and a new shallow hydrothermal vent field discovered in 2010 at a depth of 30 m, which is located close to the Faial Island, just outside the Espalamaca (38°33'N; 28°39'W)^[4]. We admitted some of the *Bacillus* strains from this collection for screening potential antimicrobial activity against pathogenic *Vibrio* and *Staphylococcus* sp.

So far conventional approaches such as use of antimicrobial drugs to control diseases have had success in the prevention or cure of disease caused by *Vibrio*, *Staphylococcus* sp. and other pathogens. The massive use of antibiotics encourages natural

emergence of antibiotic resistant bacteria, which can transfer their resistance genes to other bacteria that have never been exposed to the antibiotics. Furthermore, most of the antibiotics have been reported for the side effects for the consumers. Hence, the use of beneficial bacteria (probiotics) to displace pathogenic bacteria by competitive processes was considered as a better remedy than administering antibiotics. *Bacillus* species are used in many medical, pharmaceutical, agricultural, and other industrial processes for their ability to produce a host of enzymes, antibiotics, and other metabolites. *Bacillus* strains have also been utilized as biological controls in antibiotics and other assays. Further, several *in vivo* experimental model approaches have been used to understand the bacterial infections and drug toxicity analysis. The drug evaluation with several other mammalian models is found to be costly and slow, but zebrafish are considered for a rapid

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and high throughput analysis, which has an immune system similar to that of humans and other vertebrates including fishes. Thus, our present investigation includes, identification of potential *Bacillus* species from marine source for its ability to act as a biocontrol agent against bacterial pathogens using zebrafish as a test system.

5 selected *Bacillus* strains isolated from hydrothermal vent regions were used in this study for their antibacterial activity. *Vibrio cholera* serogroup 01 (MTCC 3904) (obtained from Microbial Type Culture Collection, IMTECH, Chandigarh, India), *V. parahaemolyticus*, *V. harveyi*, *V. alginolyticus* (isolated from fish gut, and identified using biochemical test and species specific media) and *Staphylococcus* sp. (isolated from gut of fish, grey mullet) identified using 16S rRNA sequencing by our earlier methods^[5] (obtained GenBank accession number KM029979) and *S. aureus* (obtained from Goa Medical College Hospital, Goa) strains were used for pathogens.

The screening of antagonistic activity assay was made using gram positive *Bacillus* species obtained from hydrothermal vent region against pathogens by disc diffusion method. Gram negative bacteria, *Vibrio* pathogens and *Staphylococcus* sp. were cultured in Zobell marine broth (ZMB) (HiMedia, Mumbai, India) and incubated at 28° for 24 h and 100 µl of the culture was spread over the tryptic soy agar (TSA) plates. And also TSA medium was cooled and 25 µl of pathogens were added, mixed and poured in a sterile petriplates. The disc was prepared from sterilized/autoclaved Whatman no.1 filter paper (5 mm). The 5 mm sterilized discs were placed on the cultured spread plate. Fresh culture of gram positive bacteria *B. subtilis subsp. spizizenii* and *B. thuringiensis* were centrifuged at 12 000 rpm for 10 min and the supernatant was filtered through 0.22 µm membrane filter (Millipore) for the sterile cell free extracts. Different concentrations (20, 30, 40 and 50 µl) of cell free extracts of gram positive bacteria was added into the disc of the Mueller-Hinton agar (MHA) (HiMedia, Mumbai, India) plates and incubated at 30° for 24 to 48 h. The antimicrobial activity of the isolate was said positive on any appearance of distinct inhibition zone against the pathogenic, species. Thermal stability was determined by heating a 2 ml cell-free supernatant fluid at 50°, 75° and 100° during 60 and 120 min.

The samples were then assayed for remaining activity by the well diffusion method. In order to test sensitivity

to proteases, the supernatant fluid was treated with trypsin (Sigma) and proteinase K (Sigma) at a final concentration of 0.5 mg/ml. The effect of antibacterial compound on sensitive cells was tested by adding 2.5 ml of concentrated culture supernatants of *B. subtilis subsp. spizizenii* (VSD 609) and *B. thuringiensis* (VSD 550) to 25 ml of a log phase culture of *V. cholerae* 01 and *Staphylococcus* sp. in nutrient broth. The number of viable cells determined at appropriate intervals by measuring optical density at 600 nm (OD₆₀₀) with a spectrophotometer. Ciprofloxacin/vancomycin and lysogeny broth (LB) were used as positive and negative control, respectively.

Adult zebrafish of both sexes were brought from a fish farm, Chennai, Tamil Nadu and maintained in tanks containing standard fresh water system under 14 h/10 h light-dark photo period. Fish were fed with fish food flakes thrice a day and the temperature approximately at 28°. One male and one female zebrafish were transferred to a breeding tank and separated overnight. Breeding was allowed in the next morning and the sufficient eggs were collected from the breeding tanks. The eggs were washed to remove the unfertilized eggs and debris.

Embryos were selected at the 1-cell stage and exposed 10³, 10⁴, 10⁵, 10⁶ and 10⁷ CFU/ml of bacterial pathogens in the petriplates and 6 well plates. All dechorionated embryos were observed after 24 h of incubation at 30° to monitor infection, lethality and treatment under a stereomicroscope (Olympus SZ-40, Japan) at different stages from 4 to 120 hpf and photographed for morphological analyses. The bacterial extracts wasted at different dilutions of the MIC along with bacterial pathogens for their inhibitory effect in parallel to the treatment of ciprofloxacin and vancomycin in 1% dimethyl sulphoxide (DMSO) as vehicle.

V. cholerae infection experiments using zebrafish larvae was performed as described earlier^[6]. Briefly, 5 day post-fertilization zebrafish larvae were placed into 1 ml tank water containing 1×10⁶ CFU/ml of *V. cholerae* in a 12-well plate and incubated for 2 to 24 h at 27°. At the designated time points, larvae were removed from the well with the bacteria and washed in sterile tank water twice and then placed into a well with a euthanizing dose of clove oil solution. *S. aureus* infections were made by treatment of 1×10⁵ cfu/ml of inoculum to the dechorionated 2 dpf (days post fertilization) embryos in 6 well plates incubated for 24 h^[7].

Among the five isolates of *Bacillus* species obtained

from hydrothermal vent region, *B. subtilis subsp. spizizenii* (VSD 609) and *B. thuringiensis* (VSD 550) showed antagonistic activity against *V. cholera* and *Staphylococcus* sp., respectively (fig. 1A and 1B). The level of inhibitory zone of *B. subtilis* against *V. cholerae* and *B. thuringiensis* against *Staphylococcus* sp. was consistent and distinct. The diameter of the inhibitory zone around the *V. cholerae* was about 10-15 mm and

the zone for *Staphylococcus* sp. was 14-15 mm. The cell free extracts of both the *B. subtilis subsp. spizizenii* and *B. thuringiensis* gave better zone inhibition with the concentrated supernatants when compared with that of the other live culture and diluted filtrates. Thus, the difference in the zone effect varied depending upon the concentration used (fig. 1A and 1B). The inhibitory activity was not reduced during 60 and 120 min heat

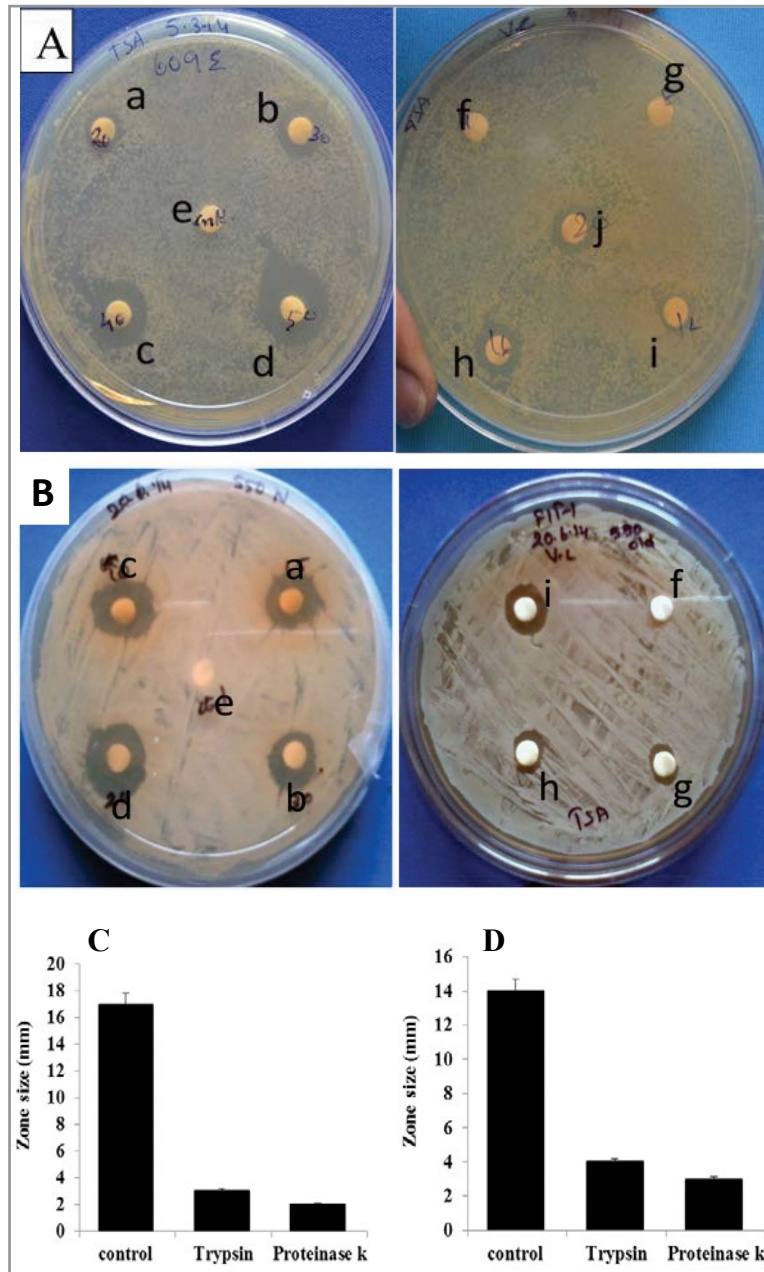


Fig. 1: Analysis of the marine *Bacillus* cell free extracts against *V. cholerae* and *S. aureus*. Inhibition of the cell free extracts of strain VSD 609, *B. subtilis subsp. spizizenii* against *V. cholerae* (A) and VSD 550, *B. thuringiensis* against *S. aureus*. (B) by the disc diffusion assay. In A, a, b, c, d and e disc containing the non-treated cell free extracts with concentrations of 20, 30, 40, 50 μ l and negative control (with only media) respectively. Af, negative control, Ag, Ah, Ai and Aj with the addition of trypsin (g,h) and proteinase k (i,j) when the antagonistic activity was completely and partially destroyed. In B, a, b, c, d and e disc containing the non-treated cell free supernatants with concentration of 20 μ l, 30 μ l, 40 μ l, 50 μ l and negative control (with only media) respectively. Bf, Bg, Bh and Bi with the addition of trypsin (f, g) and proteinase k (h, i) when the antagonistic activity was completely and partially destroyed. Zone measurements of *B. subtilis* extracts against *V. cholerae* (C) and *B. thuringiensis* against *S. aureus* (D).

treatment at 100° showed that both the supernatants of *Bacillus* sp. are heat resistant. Furthermore, the antagonistic activity showed by the supernatant fluid of both the *Bacillus* sp. was completely or partially destroyed by the trypsin and proteinase k (0.5 mg/ml, fig. 1a-e, f, g, h, i and 1b-e, f, g, h) treatment inferring that antibacterial substance was proteinaceous in nature. The spectrophotometer test on the effect of antibacterial compound of both the *B. subtilis subsp. spizizenii* and *B. thuringiensis* strains on the sensitive cells showed a stable activity in controlling the pathogens, *V. cholerae* and *Staphylococcus* sp., ranging from 0.5 to 0.8 OD after 24 h (fig. 2). The negative control in both the case showed increased population. However, bacterial pathogens treated with ciprofloxacin and vancomycin as positive controls showed significant decreased population. These results showed that the antibacterial compound exhibited the bacteriostatic mode of action.

In the present study, *B. subtilis subsp. spizizenii* showed distinct biocontrol activity against *V. cholerae* serogroup 01. *B. subtilis* is already described as an antimicrobial peptide (AMP) producer, including a diversity of lipopeptides like iturins (bacillomycins), polymyxins, fengycins (plipastatins), kurstakins^[8], and a number of bacteriocins^[9]. *B. subtilis subsp. spizizenii* also known for producing antibacterial compounds, like subtilisin A and subtilin^[10]. Similarly, *B. thuringiensis* are reported for a number of bacteriocins^[11] but only four bacteriocins have been partially characterized^[12]. Our results on antibacterial substance of heat resistant, proteinaceous nature and bacteriostatic mode of action are congruent with that of other reports^[13,14] and suggest that the antibacterial substances of the bacillus species

may be of bacteriocin related.

V. cholerae infections was made in dechorionated 2 dpf (days post-fertilization) zebrafish embryos and reisolated by placing the intestine of the larvae in the X-Gal added plates, as *V. cholerae* will form blue colonies but the other intestinal bacteria will not form blue colonies^[6,15]. The results showed that the *V. cholerae* infected larvae with more colonies when compare with that of the uninfected. Our results of *V. cholerae* infection in zebra fish larvae was found similar with that of the earlier study^[6]. The only *V. cholerae* infected embryos showed high mortality than the antibacterial treatment. The latter was treated with bacterial extracts after one hour of *V. cholerae* infection. Furthermore, the treatment of embryos with only *B. subtilis* antibacterial extracts showed no or very less *V. cholerae* colonies similar to that of the uninfected without any zebra fish larvae mortalities. Thus, our study confirms that the *B. subtilis* antibacterial extracts are safer to use as a drug.

Dechorionated embryos of 2 dpf were treated with *S. aureus* and infections were found after 24 h incubation. In order to recover bacteria from host tissues, whole zebra fish embryos were individually homogenized in a suitable volume of phosphate-buffered saline (PBS). Homogenates were serially diluted in PBS and plated on nutrient agar to determine bacterial numbers. The infected embryos were found with muscle deformities especially in trunk region and some regions like yolk sac (fig. 3). Treatment of *B. thuringiensis* substances to the infected embryos were found survived with less muscle deformities. The only *B. thuringiensis* substances treated embryos did not showed any deformities such

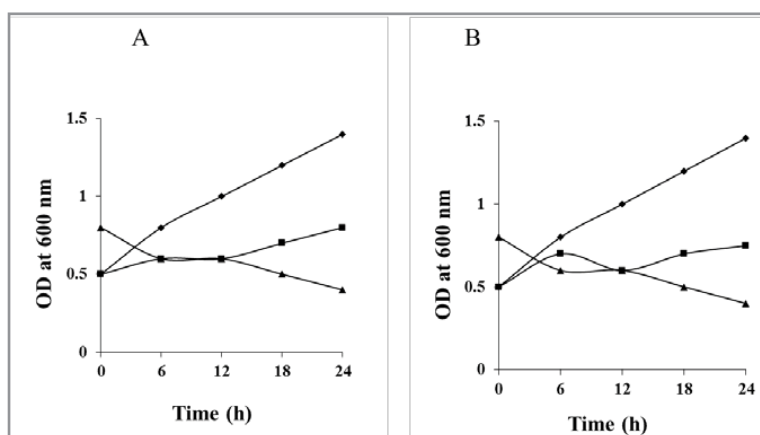


Fig. 2: Evaluation of *Bacillus* cell free extracts against the pathogens.

The inhibitory activity of inhibition of the cell free extracts of strain VSD 609, *B. subtilis subsp. spizizenii* (A) (●- Con -ve; ▲-BS; ■ Con +ve) and VSD 550 *B. thuringiensis* (B) (●- Con -ve; ▲-Bt; ■ Con +ve.) at the optimal condition against *V. cholerae*, *Staphylococcus* sp. Con +ve presents as positive control (addition of antibiotic ciprofloxacin/vancomycin in the broth culture), Con -ve presents as negative control (no addition of antibacterial in the broth culture). Values are means±SD (n=3).

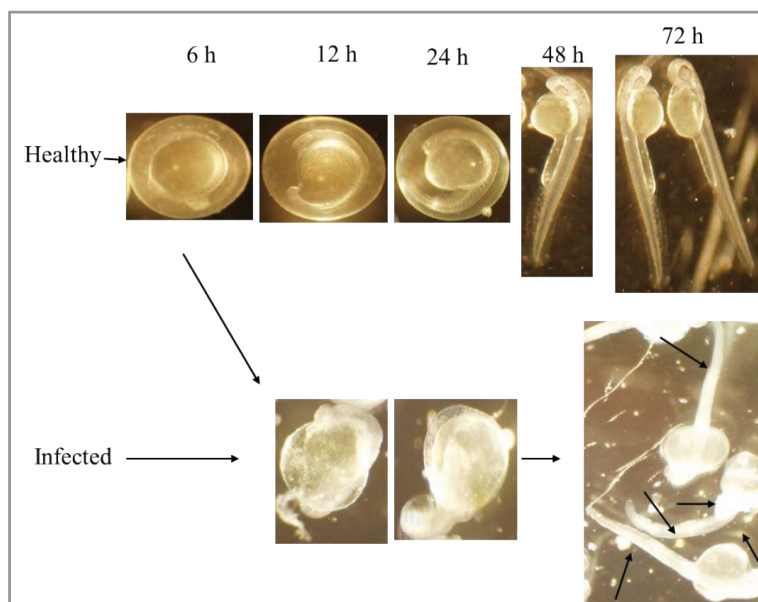


Fig. 3: Infection and treatment studies using zebra fish test system.

The toxicity and infection test was performed in the embryos. The embryos that were treated with only *Bacillus* antibacterial substances are found healthy as that of the untreated healthy embryos without any toxicity effects like muscle and other growth deformities. *S. aureus* infected embryos showed growth deformities such as muscle loss in trunk, yolk sac.

that of the vancomycin treated embryos. Thus, *Bacillus* species extracts which are found nontoxic with that of zebra fish may be tested with that of the rat models for further pathophysiological characterization against the above pathogens. Results of antibacterial substance of *B. thuringiensis* against *S. aureus* may be of bacteriocin related protein because of an earlier report stating that Gram-positive strains of bacteriocins are mostly inhibitory to Gram-positive strains^[14].

In conclusion, we elucidate the use of zebrafish, a promising model for preclinical drug assessment and toxicity assays against human pathogens. Further, the novel antibacterial substance from *Bacillus* species showed potent antibacterial activity against human pathogens without any toxicity to the host cells. Further, the antagonistic effect of the hydrothermal vent isolated *Bacillus* species may further characterized and analyzed using the rat models. This may provide valuable information for the novel drugs for therapeutic applications in controlling the infectious disease.

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Conflicts of interest:

There are no conflicts of interest.

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