A Report on the Occurrence of Thraustochytrid species in Indian Waters

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ABSTRACT...

Thraustochytrids were isolated from 52 samples of 14 substrata from the coastal waters of Goa, Tamil Nadu and Lakshadweep islands and the Arabian Sea. A total of 9 species are reported in this paper. These are: Corallochytrium limacisporum Raghukumar, Lubyrinthuloides minuta (Watson and Raper) Perkins, L. yorkensis Perkins, Thraustochytrium aggregatum Ulken, T. motivum Goldstein, T. multirudimentale Goldstein, T. striutum Schneider, Schizochytrium mangrovei Raghukumar and Ulkenia visurgensis (Ulken) Gaertner. The substrata examined were decaying mangrove leaves, living algae, estuarine, and offshore water samples, phytoplankton samples, gill tissues of green mussels, hydroid polyps, detritus samples, scrapings of oyster shell, cotal polyps and coral mucus. Ulkenia visurgensis was the most common species isolated, occurring in 57% of the substrata.

INTRODUCTION

While fungi are known to play a very important role in the terrestrial ecosystem, their function in the sea is poorly understood. The fungi-like marine protists, the thraustochytrids (Moss, 1986) are extremely common in the sea. These are unicellular, eukaryotic, osmotrophic microorganisms. Most of the reports on the occurrence of these organisms, of which about 40 species are known (Moss, 1986) have been from temperate waters. These investigations have thrown light not only on the species occurring in temperate waters but have also revealed that they occur in a wide range of substrata, Water and sediments harbour them (Bremer, 1976; Raghukumar and Gaertner, 1980) and Jacobsen, 1984) and they are associated with micro and mac oalgae (Sparrow, 1968; Gaertner, 1979; Miller and Jones, 1983 and C. Raghukumar, 1986) and invertebrates (Jones and Alderman, 1974; Hoehnk and Ulken, 1979; Wagner-Merner et al. 1980; Richter, 1985 and Alderman and Polglase, 1986). Apparently, these organisms have a definite role in the marine processes, probably similar to that of fungi in the terrestrial ecosystem. Reports of thraustochytrids from tropical waters are relatively rare (Uiken, 1984; 1986; Raghukumar, 1987 and 1988) and there is a paucity of information on the species inhabiting tropical waters and the substrata harbouring them. Such information is a necessary first step for assessing their importance in the marine ecosystem.

During ecological studies of thraustochytrids in Indian waters, many thraustochytrids were isolated from various substrata. These isolates were indentified and some results of these studies are presented here.

MATERIALS AND METHODS

The thraustochytrids were isolated either by the pine pollen baiting technique of Gaertner (1968) or by plating the substrata on a nutrient medium. Larger substrata such as algae were gently rinsed in sterile seawater prior to isolation to ensure that only cells adhering and growing on the substrata were isolated. For the baiting technique, 5 ml of sterile seawater was taken in 5 cm Petri dishes and small bits of the substratum were placed in it. A small amount

of pine pollen, sterilised in an oven at 100 C for 48 h, was sprinkled on the water. Thraustochytrids grew on the pollen in about 4-5 days. For isolating thraustochytrids from seawater, various dilutions of the water were taken in Petri dishes and baited with pine pollen as above. Such cultures also harboured bacteria. To make cultures bacteria-free, they wre grown in seawater/pine-pollen containing 5 mg streptomycin and 2000 units penicillin ml⁻¹.

In the second method, bits of the substratum were placed on a solid medium (Raghukumar, 1986) containing streptomycin and penicillin in concentrations given above. Bacteria-free colonies of thraustochytrids formed around the bits. These were subsequently subcultured onto pine pollen/seawater on which all the cultures were maintained. Observations on the morphology and development were made in paraffin chambers (Bahnweg and Sparrow, 1972) and continuous flow chambers (Raghukumar, 1987).

In situ examinations of algal bits were carried out in continuous flow chambers. Algal bits were placed in the chambers with pine pollen as the substrate. The medium was seawater containing antibiotics as above. Thraustochytrids grew under these conditions and could be identified up to the species level. The developmental patterns of all the species studied corresponded to the authentic descriptions (Goldstein, 1963 a, b; Ulken, 1965; Schneider, 1967; Perkins, 1973, 1974; Raghukumar, 1988).

RESULTS AND DISCUSSION

A list of the substrata and the thraustochytrid species which were isolated from them is presented in Table 1. A total of 9 species are reported in this paper, isolated from 52 samples of 14 different substrata. A list of the species is presented in Table 2.

Mostly only single species were isolated when the substrata were placed in petri dishes and baited or plated. However, in some cases, when the substratum was mounted directly in continuous flow chambers along with pine pollen, the presence of two or more species could be detected. Thus, filaments of the bluegreen alga, Lyngbya sp., collected from Anjuna beach, Goa, first revealed the presence of Labyrinthuloides yorkensis Perkins after 24 h in the continuous flow chamber. By 48 h, sporangia of Ulkenia visurgensis (Ulken) Gaertner were detected along with the spindle cells and ectoplasmic net system of Labyrinthuloides ininuta (Watson and Raper) Perkins. All these species completed their development in the continuous flow chamber, thus facilitating identification. Another association between Labyrinthuloides yorkensis and Elkenia visurgensis was observed when bits of the green alga Caulerpa sp. were mounted directly in continuous flow chamber and observed (Table 1).

Thus, apparently, consortia of thraustochytrid species could be present in natural substrata.

Ulkenia visurgensis was the most common species isolated, occurring in 57% of the substrata examined (Table 2). It was present in the algae Lyngbya sp. and Caulerpa sp., phytoplankton samples in the Arabian Sea, gill tissue of the green mussel, polyps of an obeliinid hydroid, water samples from the Arabian Sea, scrapings of the shells of rock oyster and in coral mucus. This species, therefore, appears to have a wide substrate range. Ulken (1984) has also found this species to be common in tropical muds. The occurrence of *U. visurgensis* in hydroid polyps has been reported in detail elsewhere (Raghukumar, 1988). The isolation of this species from the gill tissue of the green mussel might be related to the filter feeding

TABLE 1
List of thraustochytrid species and their substrata

1. Decaying mangrove leaves (a) Khizophora murconata Lam. (b) Avicennia officinalis L. (c) Avicennia officinalis L. (b) Avicennia officinalis L. (c) Avicennia officinalis L. (d) Avicennia officinalis L. (e) Avicennia officinalis L. (f) Avicennia officinalis L. (g) Avicennia officinalis L. (h) Avicennia L. (Substratum	Date and Locality (No. of samples)	Species
(b) Avicennia officinalis L. Blue-green alga, Lyngbya sp. Brown-alga, Sargassum sp. Green-alga Caulerpa sp. Red-alga Laurencia sp. Estuarine water sample Study goa, Dec. 1982 (1) Estuarine water sample Study goa, Dec. 1982 (1) Estuarine water sample Central Arabian Sea, Sagar Kanya Cruise I, July 1983 (2) Gill tissue of the green mussel Phytoplankton eamples Offshore water samples Offshore water sample	 -	Decaying mangrove leaves (a) Rhizophora mucronata Lam.	Pitchavaram mangroves, Tamil Nadu, June 1982 (1) Orda, Goa, December 1982 (1) Orda, Goa, April 1986 (4)	Thraustochytrium striatum Schneider T. striatum Schneider T. motivum Goldstein T. aggregatum Ulken Schizochytrium mangrovei Raghukumar
Brown-alga, Lyngbya sp. Brown-alga, Sargassum sp. Green-alga Caulerpa sp. Red-alga Laurencia sp. Estuarine water samples Phytoplankton samples Phytoplankton samples Polyps of an obeliinid hydroid Polyps of an obeliinid hydroid Detrital particles from an intertidal pool Shells of the rock oyster Mucus from the stag-horn coral Arabian beach, Goa, June 1987 (3) Mandapam, Tamil Nadu, March 1986 (1) Mandapam, Tamil Nadu, April 1986 (1) Zuari estuary, Goa, Dec. 1982 (1) Zuari estuary, Goa, Dec. 1982 (1) July 1983 (2) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Detrital particles from an intertidal pool Anjuna beach, Goa, April 1986 (2) Anjuna beach, Goa, April 1986 (1) Lakshadweep islands, Minicoy (3) Lakshadweep islands, Minicoy (3)		(b) Avicennia officinal is L.	Orda, Goa, May 1987 (1)	S. mangrovei Raghukumar Thraustochytrium nultirudimentale Goldstein
Green-alga, Sargassum sp. Green-alga Caulerpa sp. Red-alga Laurencia sp. Estuarine water samples Phytoplankton samples Phytoplank	6		Anjuna beach, Goa, June 1986 (1)	Labyrinthuloides yorkensis Perkins L. minuta (Watson and Raper) Perkins Ulkenia visurgensis (Ulken) Gaertner
Green-alga Caulerpa sp. Red-alga Laurencia sp. Estuarine water sample Phytoplankton samples Gill tissue of the green mussel Polyps of an obeliinid hydroid Offshore water samples Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mandapam, Tamil Nadu, March 1986 (1) Zuari estuary, Goa, Dec. 1982 (1) Zuari estuary, Goa, Dec. 1982 (1) Arambol beach, Goa, October 1986 (1) Baga beach, Goa, March 1986 (5) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Arabian beach, Goa, April 1986 (2) Shells of the rock oyster Corals (Acropora spp.) Lakshadweep islands (21) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)			Anjuna beach, Goa, June 1987 (3)	Labyrinthuloides minuta (Watson and Raper) Perkins
Red-alga Laurencia sp. Estuarine water sample Phytoplankton samples Gill tissue of the green mussel Polyps of an obeliinid hydroid Offshore water samples Offshore water samples Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Eakshadweep islands (21) Mandapam, Tamil Nadu, April 1986 (1) Central Arabian Sea, Sagar Kanya Cruise 1, July 1983 (2) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Arabian beach, Goa, April 1986 (1) Lakshadweep islands (21) Mucus from the stag-horn coral Eakshadweep islands, Minicoy (3)	4.		Mandapam, Tamil Nadu, March 1986 (1)	L. yorkensis Perkins Ulkenia visurgensis (Ulken) Gaertner
Estuarine water sample Phytoplankton samples Central Arabian Sea, Sagar Kanya Cruise 1, July 1983 (2) Gill tissue of the green mussel Polyps of an obeliinid hydroid Offshore water samples Offshore water samples Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Lakshadweep islands, Minicoy (3) Estuarios Coa, Dec. 1982 (1) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Anjuna beach, Goa, April 1986 (2) Anjuna beach, Goa, November 1986 (1) Lakshadweep islands (21) Lakshadweep islands, Minicoy (3)	s,		Mandapam, Tamil Nadu, April 1986 (1)	Thraustochytrium motivum Goldstein
Phytoplankton samples Gill tissue of the green mussel Polyps of an obelinid hydroid Offshore water samples Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Central Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Anjuna beach, Goa, April 1986 (2) Anjuna beach, Goa, November 1986 (1) Lakshadweep islands (21) Lakshadweep islands, Minicoy (3)	6		Zuari estuary, Goa, Dec. 1982 (1)	T. striatum Schneider
Gill tissue of the green mussel Polyps of an obelinid hydroid Polyps of an obelinid hydroid Offshore water samples Offshore water samples Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	7.	Phytoplankton samples	Central Arabian Sea, Sagar Kanya Cruise 1, July 1983 (2)	Ulkenia visurgensis (Ulken) Gaertner
Polyps of an obeliinid hydroid Offshore water samples Offshore water samples Offshore water samples July 1983 (3) Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	œ.		Arambol beach, Goa, October 1986 (1)	U. visurgensis (Ulken) Gaertner
Offshore water samples July 1983 (3) Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3) Anjuna beach, Goa, April 1986 (2) Anjuna beach, Goa, November 1986 (1) Lakshadweep islands (21) Lakshadweep islands, Minicoy (3)	ģ		Baga beach, Goa, March 1986 (5)	U. visurgensis (Ulken) Gaertner
Detrital particles from an intertidal pool Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	10.		Arabian Sea, Sagar Kanya Cruise 1, July 1983 (3)	U. visurgensis (Ulken) Gaertner
Shells of the rock oyster Corals (Acropora spp.) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	=		Anjuna beach, Goa, April 1986 (2)	Schizochytrium mangrovei Raghukumar
Corals (Acropora spp.) Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	12.		Anjuna beach, Goa, November 1986 (1)	Ulkenia visurgensis (Ulken) Gaertner
Mucus from the stag-horn coral Lakshadweep islands, Minicoy (3)	13.		Lakshadweep islands (21)	Corallochytrium limacisporum Raghukumar
	4.		Lakshadweep islands, Minicoy (3)	C. Ilmacisporum Raghukumar Ulkenia visurgensis (Ulken) Gaertner Labyrinthuloides yorkensis Perkins

TABLE 2
Percentage of substrata harburing the various species of thraustochytrids

	Species	Percentage
1.	Corallochytrium limacisporum Raghukumar	14.3
2.	Labyrinthuloides minuta (Watson and Raper) Perkins	14.3
3.	L. yorkensis Perkins	21.4
4.	Schizochytrium mangrovei Raghukumar	14.3
5.	Thraustochytrium aggregatum Ulken	7.1
6.	T. motivum Goldstein	14.3
7.	T. multirudimentale Goldstein	7.1
8.	T. striatum Schneider	14.3
9.	Ulkenia visurgensis (Ulken) Gaertner	57.1

habits of the animal, resulting in the concentration of cells of this thraustochytrid from water in the gill.

Labyrinthuloides yorkensis was isolated from the algae Lyngbya sp. and Caulerpa sp. and from the mucus of the staghorn coral in the Lakshadweep islands. This species has also been reported by the author from Central Arabian Sea (Raghukumar, 1986) and therefore appears to be quite common. It had a percentage occurrence on 21.4 substrata examined (Table 2). The results also suggest that some species have a narrow substrate range (Tables 1 and 2). Corallochytrium limacisporum Raghukumar was isolated only from the coral reef lagoon of Lakshadweep islands, in coral polyps, mucus and lagoon waters. This species is very comon in the lagoon waters of Lakshadweep islands (Raghukumar, 1987a), but was not isolated from any substratum outside it. Thraustochytrium aggregatum Ulken and T. multirudimentale Goldstein occurred in only 7.1% of the substrata examined and were isolated only once from decaying leaves of the mangrove, Rhizophora mucronata Lam.

Only qualitative aspects are presented in this paper. Quantification of thraustochytrids on the various substrata is needed in future to study their ecology Raghukumar (1988) demonstrated Ulkenia visurgensis in great numbers in an obeliinid hydroid, using immunofluorescence and presented some data on their numbers in the waters of the Arabian Sea (Raghukumar, 1986). These aspects will have to be studied in greater detail in future, using techniques which are availabe for such quantitative studies (Gaertner, 1968; Raghukumar, 1986).

REFERENCES

ALDERMAN, D. J. AND POLGLASE, J. L. 1986. Are fungal diseases significant in the marine environment? In: MOSS, S.T. (ed.) The Biology of Marine Fungi, (pp. 190-198). Cambridge University Press, Cambridge.

Bahnweg, G. and Sparrow, F. K. JR. 1972. Aplanochytrium kerguelensis gen. nov. spec. nov., a new Phycomycete from Subantarctic marine waters. Arch. Mikrobiol. 81: 45-49.

BREMER, G. B. 1976. The ecology of marine lower fungi. In: Gareth Jones, E. B. (ed.) Recent advances in aquatic mycology. (pp. 313-333). Elek Science, London.

- GAERTNER, A. 1968. Eine Methode des quantitativen Nachweises niederer, mit Pollen Koederbarer Pilze im Meerwasser und im Sediment. Veroeff. Inst. Meeresforsch. Bremerh. 3: 75-91.
- GAERTNER, A. 1979. Some fungal parasites found in the diatom populations of the Rosfjord area (South Norway) during March 1979. Veroeff. Inst. Meeresforsch. Bremerh. 18: 29-33.
- GOLDSTEIN, S. 1963a. Development and nutrition of a new species of *Thraustochytrium*. Amer. J. Bot. 50: 271-279.
- GOLDSTEIN, S. 1963b. Studies of a new species of Thraustochytrium that displays light stimulated growth. Mycologia 55: 799-811.
- HOEHNK, W. AND ULKEN, A. 1979. Pilze aus marinen Schwaemmen. Veroeff. Instit. Meeresforsch. Bremerh. 17: 99-204.
- JACOBSEN, K. R. 1983. Thraustochytriaceae im Interstitium ausgewaehlter Sandwatten des Weseraestuars. Veroeff. Inst. Meeresforsch. Bremerh. 19:145-175.
- Jones, E.B.G. and Alderman, D. J. 1974. Althornia crouchii gen. et sp. nov., a marine biflagellate fungus. Nova Hedwigia 21: 381-399.
- MILLER, J. AND JONES, E.B.G. 1983. Observations on the association of thraustochytrid marine fungi with decaying seaweed. Botanica Marina 26:345-351.
- Moss, S. T. 1986. Biology and phylogeny of the Labyrinthulales and Thraustochytriales. In: Moss, S.T. (ed.) The Biology of Marine Fungi. (pp. 105—130). Cambridge University Press, Cambridge.
- Perkins F. O. 1973. A new species of marine labyrinthulid *Labyrinthuloides yorkensis* gen. nov. spec. nov.—Cytology and fine structure. Arch. Mikrobiol. 90: 1-17.
- Perkins, F. O. 1974. Reassignment of Labyrinthula minuta to the genus Labyrinthuloides. Mycologia 66: 697-702.
- RAGHUKUMAR, C. 1986. Thraustochytrid fungi associated with marine algae. Indian J. mar. Sci. 15: 121-122.
- RAGHUKUMAR, S. 1986. Enumeration of thraustochytrds (heterotrophic microorganisms) from the

- Arabian Sea, Mahasagar, Bull. natl. Inst. Oceanogr. 18: 457-465.
- RAGHUKUMAR, S. 1987a. Occurrence of the thraustochytrid Corallochytrium limacisporum gen. et sp. nov. in the coral reef lagoons of the Lakshadweep islands in the Arabian Sea. Botanica Marina 30: 83-89.
- RAGHUKUMAR, S. 1987b. A device for continuous microscopic observations of aquatic microorganisms. Indian J. mar. Sci. 16: 132-133.
- RAGHUKUMAR, S. 1988a. Schizochytrium mangrovei sp. nov.: a thraustochytrid from mangroves in India. Trans. Br. mycol. Soc. 90: 627-631.
- RAGHUKUMAR, S. 1988b. Detection of a thraustochytrid protist in a hydroid, using immunofluorescence. Mar. Biol. 97: 253-257.
- RAGHUKUMAR, S. AND GAERTNER, A. 1980. Ecology of the thraustochytrids (lower marine fungi) in the Fladen Ground and other parts of the North Sea. U. Veroeff. Inst. Meeresforsch. Bremerh. 18: 289-308.
- RICHTER, W. 1985. Marine Schwaemme als Substrat fuer Thraustochytriaceae (marine niedere Pilze). Veroeff. Inst. Meeresforcsh. Bremerh. 20:141—149.
- Schneider J. 1967. Ein neuer Phycomycet aus der Kieler Bucht (Thraustochytrium striatum spec. nov.) Kieler Meeresforsch. 23: 16-20.
- Sparrow, F. K. Jr. 1968. On the occurrence of the Thraustochytriaceae. Veroeff. Inst. Meeresforsch. Bremerh. 11: 89-92.
- ULKEN, A. 1965. Zwei neue Thraustochytrien aus der Aussenweser. Veroeff. Inst. Meeresforsch. Bremerh. 9: 289-295.
- ULKEN, A 1984. The fungi of the mangal ecosystem. In: Por, F. D. and Dor, I. (eds.) Hydrobiology of the mangal (pp. 27-33). Dr. W.Junk Publishers, The Hague.
- ULKEN, A 1986. Estimation of thraustochytrid propagules in two mangrove swamps, Botanica Marina 29: 85-89.
- WAGNER-MERNER, E., DUNCAN, W. R. AND LAWRENCE, J. M. 1980. Preliminary comparison of Thraustochytriaceae in the guts of a regular and irregular echinoid. Botanica Marina 23: 95-97.