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Dinoflagellates cyst assemblage concerning trophic index for eutrophication from major ports along the west coast of India

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Abstract:

An overview of dinoflagellates cysts assemblage is presented as a trophic index for three monsooninfluenced estuarine and marine ports along the Indian coast. The cyst distribution (including harmful species) showed a trend of highest abundance and species number in highly eutrophicated estuarine (Cochin-south) followed by medium (New-Mangalore-central) and low (Kandla-north) levels of eutrophicated marine ports. The investigation revealed four new species in the region (*Bitectatodinium spongium, Gonyaulax elongatum, Brigantedinium* sp. and potential harmful species *Blixaea quinquecornis*-cyst similar to planktonic). Autotrophs dominance in the highly productive Cochin and New-Mangalore ports reveals that, in eutrophic systems, heterotrophs need not always be dominant. The indicator taxa (*Polykrikos, Protoperidnium*, and *Lingulodinium*) presence in high density indicated a eutrophic system. This study concludes cyst (species numbers/Fisher- α index/indicator species) as potential eutrophication proxies and emphasizes greater harmful-algalbloom risks in the high trophic-index ports (Cochin and New-Mangalore).

Keywords: Dinoflagellate cyst, ports, eutrophication, trophic index, Harmful Algal Blooms.

Introduction

Dinoflagellates are the most dominant and diverse phytoplankton group in freshwater, estuarine, and marine environments and also include potential invaders. Cysts production, as part of many dinoflagellate life cycles has been reported. In some cases, cysts are produced in response to adverse environmental conditions, for example, changes in temperature, salinity, and nutrient fluxes. Once formed, most of the resting cyst settles on the sediment bottom. They remain viable for several years and acts as an essential source of the planktonic population when suitable conditions return (Matsuoka and Fukuyo 2000). Resting stages are referred to as "time capsules," as they preserve biological material for a long time (Ribeiro et al., 2013). In recent years, concerned with the global increase in the occurrence of Harmful Algal Blooms (HABs) in coastal waters, owing to eutrophication (anthropogenic nutrient enrichment; e.g. Anderson et al., 2002), changing climate patterns (eg. Hallegraeff, 2010) and ships ballast water-mediated introduction of non-native species (eg. Smayda, 2007; Casas-Monroy, et al., 2013), the studies on dinoflagellates and its cysts have received importance. As several of the dinoflagellate species are responsible for the occurrence of HABs worldwide, some of which are toxin-producing and act as environmental proxies. At least 90 of these cysts producing species are known to cause harmful blooms (Sournia et al. 1995), and 45 species are considered toxic (Sournia et al. 1995; Smayda et al. 1997; Hallegraeff et al. 2003). Further, dinoflagellate cyst assemblages are also considered as a valuable tool (for the detection and mapping of HABs; Satta et al. 2013) and proxy to determining past environmental variations in the water conditions (such as changes in sea-surface temperature, salinity, turbulence, and eutrophication along with changes in sedimentary conditions; Zonneveld et al. 2012).

A global atlas on cysts distribution studies in the world ocean suggests that a considerable amount of information is available from all the climatic zones of the world oceans except from the Indian subcontinent (Zonneveld et al., 2013). Despite the above-said importance, the information on dinoflagellate cysts distribution from the Indian subcontinent is scarce and are restricted to modern cysts at few locations (D'Costa et al., 2008; D'Silva et al., 2012, 2013; Godhe et al. 2000; Patil 2003; Narale et al., 2013, 2015, Narale and Anil, 2017) as well as from the sediments representing Late Quaternary period from Bay of Bengal (Naidu et al. 2012). Considering India being one of the significant maritime countries and having a vast coastline (~7,500 km), the studies on cysts producing dinoflagellate need detailed investigation at spatial and temporal scales to better understand their population dynamics holistically. Moreover, Indian coasts are characterized with high particulate-load and high species diversity but low cyst abundance. Therefore characterizing the distributional patterns of dinoflagellate cysts will be beneficial in improving the understanding of HABs and also to study the spatio-temporal change of dinoflagellate cysts in different environments.

Considering human induced alterations over the years, it is presumed that the influence of such alterations (including eutrophication) will be reflected in dinoflagellates and cyst assemblages *per se*. In this study, the information (such as composition and species diversity and richness) on cyst assemblages from three major ports (Cochin – estuarine Port, Mangalore – marine port, and Kandla – marine port experiencing hypersaline condition during non-monsoon) located along the west coast of India was correlated with the trophic status (by comparing with available indices for eutrophication/pollution) for identifying new environmental proxies. The data generated will provide additional information to the growing body of knowledge on dinoflagellates as water quality proxies and biogeography, including harmful and indicator (nutrient enrichment) species for the less studied monsoon influence tropical ports. The sampling in each port was carried out as part of the port biological baseline survey for BAMPI (Ballast water Management Program, India).

Materials and methods

Description of the study area

Cochin Port

Cochin port (Kerala, India) is one of the major Indian ports and is situated on a bar-built estuary at the convergence of backwaters. Cochin ports, Ernakulum channel is 5km long and 200 mts wide, while Mattancherry channel is 2.6 km long and 180 m wide (BAMP Monograph series No. 5, Internal publication). Cochin receives 55–65 % of the total annual rainfall during the monsoon season (Menon et al., 2000). The salinity of the backwaters reaches near-zero values due to the heavy precipitation during the peak monsoon period. The maximum tidal range is 1 m in the port regions, whereas the influence of tides is insignificant (Qasim 2003).

For sampling, a total of 23 stations were selected based on activities such as fishing harbor, jetties, wharf, dry docks, creek mouth, and berths (Fig.1 and suppl. Table 1). Sampling was carried out on four occasions representing different seasons, i.e., post-monsoon (October 2011 and November 2012), pre-monsoon (May 2012), and August 2012 (monsoon). The sampling was performed from morning till noon and was spread over 5 to 6 days on each occasion.

New Mangalore port

New Mangalore Port (Karnataka) is a modern all-weather on the West Coast of India. New Mangalore ports, side arm channel is 900 mts long and 198 mts wide, Oil berth channel is 1055 m

long and 234 m wide (BAMP Monograph series No. 9, Internal publication). Iron ore pellets and concentrates, petroleum, oil and lubricants (POL) products, granite stones, container cargo, etc. are the major commodities exported from this port. On the other hand, the major imports of the Port are POL products and crude oil, LPG, Coal, Limestone, Finished Fertilizers, Liquid Ammonia, Phosphoric Acid, Other Liquid Chemicals, Containerized cargo, etc.

Sampling was carried out on four occasions representing different seasons, i.e., post-monsoon (November 2011 and December 2012), pre-monsoon (May 2012), and monsoon (September 2012) in New Mangalore Port. In this study, the water and sediment samples were sampled from 19 stations based on port activities (Fig.1 and suppl. Table 2) for dinoflagellate motile-stages and cysts, respectively. Tides ranged from 0.4 to 1.2 m during the study period.

Kandla Port

Kandla Port, which is located along the western bank of Kandla creek, is operational throughout the year and thereby referred to as an all-weather port. Kandla ports, channel width vary from 200 to 1000 m and total length is around 28 kms (BAMP Monograph series No. 7, internal publication). Various fertilizer-manufacturing industry, industrial-chemical manufacturing units, salt manufacturing units with saltpans rich in brines are located around the Kandla creek (Shirodkar et al. 2010). The port facilitates the extensive traffic of passenger cargo vessels, ore carriers, oil tankers, freighters, fishing boats, and container vessels in Kandla port. In this study, a total of 26 stations were selected based on port activities (Fig.1 and suppl. Table 3), and sampling was done on four occasions in post-monsoon (October 2014 and 2015), monsoon (August 2015), and pre-monsoon (February 2016).

Sampling

5-L Niskin sampler was used to collect surface and near-bottom water samples. A multiparameter portable instrument (SONDE) was used to measure temperature and salinity at the site. Dissolved oxygen, biological oxygen demand, and chlorophyll-*a* were estimated by following standard procedures (Parsons, 2013). Nutrient concentrations such as phosphate, silicate, nitrite, nitrate, and ammonia were determined using an autoanalyzer (Skalar San++). To evaluate the trophic status of port waters, TRIX was calculated using the equation TRIX= (log10 (chl *a* × a%O₂ × DIN × DIP) - k)/m (Vollenweider et al. 1998). TRIX scores lesser than 3-4 indicate low eutrophication; scores of 4-5 indicate medium eutrophication with; scores of 5-6 indicate high eutrophication, and scores > 6 indicate elevated levels of eutrophication. Trix score was used to assess the relationship with the dinoflagellates cyst. The Vollenweider 1998 TRIX eutrophication score have been successfully

implemented in Mediterranean ports (Massi et al. 2019) and was also employed in other regions, including ports (eg. Rajaneesh et al 2015, Noyel and Desai, 2020). In this study the sampling was conducted continuously over 3-4 days encompassing different tidal regimes and the TRIX score during this sampling period don't vary significantly (for either of the three ports) suggesting negligible influence of the physical forcing.

Van Veen grab was used to collect bottom sediment samples with a 0.04 m^2 grabbing area. In addition, undisturbed sediment cores (20 cm in length PVC corer with an inner diameter of 2.5 cm) through the top windows of the grab, were collected. Cores were sealed with caps, kept in ice in the dark containers. In the laboratory, sediment cores were sectioned at 2 cm interval, mixed well, and stored in airtight plastic bags at 4 0 C in the dark until further dinoflagellate cyst analysis.

Dinoflagellate cyst analysis

Only the top fraction (0–2 cm), which comprises the recent sampled sediments, was processed using the palynological method (Matsuoka and Fukuyo 2000). Sediment samples were sonicated for 10 mins and sieved through 100 and 10 μ m mesh-sizes. The residue retained on the 10 μ m mesh was acid-treated with 10% HCL acid and 30% HF acid with distilled water washes in between to remove the acid. Next, the sample was sieved through 10 μ m mesh size. The residue retained on the mesh was transferred and suspended in 10 mL of distilled water into a vial. Aliquots of 0.25–1 mL of the residual samples were diluted to a total volume of 2.5 mL in a Petri dish (diameter 3.8 cm) and scanned under an inverted microscope (Olympus IX 71) at 100x to 400x magnification. Dinoflagellate cysts were identified based on published literature (Wall and Dale 1968; Sonneman and Hill 1997; Matsuoka and Fukuyo 2000). The cyst abundance was expressed as the number of cysts g⁻¹ dry sediment after calculating the water content according to the formula given by Matsuoka and Fukuyo (2000).

Results

Environment

In all the studied ports, the measured environmental parameters revealed a distinct seasonal variation (for details, please see Rath et al. 2018 for New Mangalore port, Rodrigues et al. 2019 for Cochin port, Sathish et al. 2020 for all three ports). However, the seasonal trends between the ports were different (Rath et al. 2018, Rodrigues et al. 2019, Sathish et al. 2020.) The averaged salinity variations were small (30–40 psu) in marine ports (Kandla, New Mangalore), and large (8–38 psu) estuarine ports (Cochin). The low salinity observed in marine and estuarine ports was due to riverine discharge and precipitation during monsoons. Water temperatures in all three ports are relatively

warmer in pre-monsoon and cooler during southwest monsoon (SWM) and post-monsoon. Interestingly, nutrients (NO3, PO4, and SiO4) were more in estuarine than marine port systems. However, the concentrations of nutrients varied between the ports. The nutrient concentrations were higher for Kandla as compared to Cochin and New Mangalore ports (Table 1). Nutrients showed a distinct seasonal variability among the marine ports, but the seasonal trends varied between the ports located. The trophic index (TRIX) revealed that Kandla port (3-4) had a higher state of water quality with low eutrophication level, Mangalore port (4-5) had a good state of water quality with medium eutrophication level, and Cochin port (5-6) had a bad state of water quality with high eutrophication level.

Dinocyst distribution, and diversity

Overall a total of 40 dinocysts were reported belonging to 17 genera and 2 unidentified dinocysts in the three different port ecosystems (Table 1; Figs. 2 and 3). The autotrophic and heterotrophic dinocyst consists of 20 species each. In addition, a total of 8 potentially HAB species were also recorded in this study. The distribution of dinocyst abundance showed distinct variations between the ports. Dinocyst abundance was highest in Cochin station (1721 cyst⁻¹g of DW sediment), followed by Mangalore station (864 cyst⁻¹g of DW sediment) and Kandla station (24 cyst⁻¹g of DW sediment). This study revealed a high species number in Cochin (35 species) followed by New Mangalore (32 species) and Kandla (15 species) (Fig. 3 b). A total of 7, 8, and 3 species found in Cochin, New Mangalore, and Kandla port are known to be potential HAB species (Table. 2). Of the 8 HAB species, 7 were autotrophic forms, and one was heterotrophic form. The common HAB species found in all ports were Gonyaulax spinifera complex, Lingulodinium polyedra, and Protoceratium reticulatum. In New Mangalore and Cochin port, HAB species found were Alexandrium tamarense complex, Cochlodinium sp, Gonyaulax scrippsae, and Scrippsiella acuminata (Table.2). In contrast, HAB species of Blixaea quinquecornis was found only in New Mangalore port. The highest HAB dinocyst abundance was recorded in Cochin, followed by New Mangalore and Kandla port. Overall the autotrophic dinoflagellates dominated in Cochin and New Mangalore ports (Fig. 3b). While heterotrophic dinoflagellates were dominated in Kandla port (Fig. 4b). Pyrophacus steinii and Gonyaulax spinifera complex were the most dominant species in Cochin and New Mangalore ports (Fig. 3a). Brigantedinium spp and Bitectodinium sponginum were the dominant species in the Kandla port (Fig. 3b).

The univariate measures, such as species number, species richness, species diversity, and fisher's index, were also highest in Cochin, followed by New Mangalore and then in Kandla port (Fig. 4a, c, d, e, and f). The ANOVA of dinocyst abundance, species richness, species diversity,

fisher's index, and species number showed a significant variation among the ports. The comparison of the ports revealed a total of 9 species, which were common between the three ports, of which 5 are autotrophic, and four are heterotrophic forms. Of these 9 species, 4 species (Gonyaulax spinifera complex, Lingulodinium polyedra, Protoperidinium pentagonum, and P. leonis) were some of the dominant species in each of these ports (Fig. 3). A total 5 species (Bitectatodinium spongium, Brigantedinium spp., Gymnodinium microreticulatum. Protoperidinium avellana. and Protoperidinium denticulatum) were restricted to only to Kandla port. In comparison, a total 3 species (Pyrodinium sp1, Pyrophacus sp, and Protoperidinium sp1) and 1 species (Blixaea quinquecornis) were restricted to Cochin and New Mangalore ports, respectively. Cluster analysis of the dinocyst abundance showed that Cochin and New Mangalore ports were similar to Kandla port (Fig. 6a) as these ports have identical species composition. Cluster analysis of species showed grouping, and at 50% similarity, it formed 2 groups (Fig. 6b). Group, I consist of 7 species represented by rare species with low abundance during the study period (Fig. 6b). While Group II comprised 32 species and was again clustered into two subgroups (IIa, consist of 28 species, and IIb, consisting of 4 species). The species in subgroup IIa, were the dominant species, while species representing subgroup II b are rare species with higher abundance (Fig. 6b).

Relationship between the community indices and trophic index (TRIX)

The community indices (abundance, species number, diversity, and richness and fisher's index) versus the TRIX score indicated different scenarios between the ports. In Kandla port, the community indices were the lowest along with low eutrophication levels compared to the other two ports (Fig. 5). In New Mangalore port, the community indices were intermediate along with medium eutrophication levels as compared to the other two ports (Fig. 5). In Cochin port, the community indices were higher along with high eutrophication levels than the other two ports (Fig. 5). In Cochin and New Mangalore Port, the autotrophic cyst abundance was higher with high TRIX scores as compared to Kandla port (Fig. 5a). In Cochin, New Mangalore, and Kandla port, the heterotrophic dinocyst species number and species diversity were higher with a high TRIX score (Fig. 5a). In Cochin and New Mangalore port, the heterotrophic cyst fisher's index was higher with high TRIX scores (Fig. 5a). In Cochin and New Mangalore port, the heterotrophic cyst fisher's index was higher with high TRIX score (Fig. 5a). In Cochin and New Mangalore port, the heterotrophic cyst fisher's index was higher with high TRIX scores as compared to Kandla port (Fig. 5d). In Cochin and New Mangalore port, the autotrophic cyst fisher's index was higher with high TRIX scores as compared to Kandla port (Fig. 5d). In Cochin and New Mangalore port, the autotrophic cyst fisher's index was higher with high TRIX scores as compared to Kandla port (Fig. 5d). In Cochin and New Mangalore port, the autotrophic cyst species richness was high at higher TRIX score, and the heterotrophic cyst was high at low TRIX score in Kandla (Fig. 5e).

Discussion

Dinocyst assemblage diversity and distribution

The present survey on dinoflagellate cyst diversity from three major ports, with different environmental settings along the west coast of India, revealed 42 species. The highest number of cyst types and abundance was recorded in Cochin (estuarine port), followed by New Mangalore port and the least in Kandla (marine but hypersaline) port (Fig. 3a). The prevalence of silty sediment round the year in the Cochin and New Mangalore port could be the reason for the high cyst abundance compared to Kandla ports (Table. 1). Studies suggest that the higher cysts abundances are directly proportional to higher dinoflagellate production in areas with fine-grained sediments (Dale 1983; Anderson et al. 1995; Joyce et al. 2005; Rodrigues et al. 2019). Even considering the reported cyst assemblage from the other major ports Mumbai and JNPT Ports (D'Costa et al. 2008) and Goa (D'Silva et al. 2012) along the west coast of India, cyst abundance and diversity was highest and lowest in Cochin and Kandla port, respectively (Table. 2). These findings suggested that the dinoflagellate cyst abundance and species number showed a decreasing trend with respect to the geographical location of the ports (i.e., from south to north) in the following order: Cochin (southern port) - New Mangalore - Goa - Mumbai - Kandla (northern port). Interestingly, the change in autotrophic and heterotrophic dominance according to the geographical location of the port was also evident. For instance autotrophic and heterotrophic cysts were dominant in southern (Cochin and New Mangalore) and northern (Mumbai/JNPT and Kandla) ports, respectively in the present study. D'Silva et al. (2012) reported heterotrophic dominance along the west coast of India. Still, Rodrigues et al. (2019) observed a decadal change in the dominance from heterotrophic to autotrophic forms in the Cochin Port. Such a paradigm shift might be due to change in environmental settings as the autotrophs and heterotrophs respond directly and indirectly to the environment.

Species composition revealed that though most of the species were recorded in the studied ports, few species with port-specific distribution are considered uncommon species. For instance, the species number of 5, 1, and 3 occurred only in Kandla, New Mangalore, and Cochin Port, respectively (Fig. 3). Before this study, available reports from the major ports (Goa, Mumbai, and Mangalore), suggested the presence of 69 cysts types (Godhe et al. 2000; D'Costa et al. 2008; D'Silva et al. 2012, 2013), and with the present study, due to record of 6 new species records (including one potential HAB species), the total no of cysts types is 75. The four new species include *Bitectatodinium spongium* (autotrophic), *Gonyaulax elongatum* (autotrophic), *Blixaea quinquecornis* (heterotrophic and potential HAB species), *Brigantedinium* spp. (heterotrophic). Among the studied ports, only Cochin port has some cyst data for the year 2000 (D'Silva et al. 2012), and the

comparison indicated that the present study listed 11 more cyst types (including three potentially harmful). Even the density of the dominant species (some *Protoperidinium* spp. and *Gonyaulax spinifera* complex) showed an increasing trend over the last decade (Rodrigues et al. 2019). Nevertheless, the detailed checklist of cyst types and morphological details presented here as part of the port biological baseline survey (PBBS) will be valuable in the studies related to microalgal (dinoflagellate per se) biodiversity and invasion, hereafter.

Documenting cysts of harmful dinoflagellates from the field is considered one of HAB monitoring strategies (Anderson et al., 2012; Ribeiro et al., 2012) as viable cysts can be a precursor for blooms. Given this, the present study lists potential 11 HAB species from the six major ports along the west coast of India (Table 2). Of the 11 species, 9 (Alexandrium tamarense complex, Cochlodinium sp, Gonyaulax scrippsae, G. spinifera complex, Scrippsiella acuminata, Protoceratium reticulatum, Lingulodinium polyedra, Pyrodinium bahamense and Blixaea quinquecornis) potentially HAB species are documented, highlighting the potential HAB risks under favorable conditions. Most of these HAB species are known to produce various toxins (e.g. icthyotoxins, saxitoxins, and yessotoxins), and also responsible for oxygen depletion and discoloration of water (Table 2). The following dinoflagellates, Alexandrium tamarense complex, Scrippsiella acuminata, Gonyaulax spinifera complex, Protoceratium reticulatum, and Lingulodinium polyedra, which are known for triggering potentially toxic HABs, and these species were most dominant in the Cochin and New Mangalore ports (Fig. 3). Although no major HABs have been reported during the investigation, the previous HAB instances or shellfish poisoning outbreaks in and around the Cochin and New Mangalore ports have also been documented (Madhu et al., 2011; Padmakumar et al., 2012, 2018). Cysts of saxitoxin producing species Pyrodinium bahamense have also been reported from Mumbai port on west coast (Table 3) and Visakhapatnam harbour along the east coast of India (D'Silva et al. 2013). However, to our knowledge reports of its bloom are reported. Recent reports of Gonyaulax spp. blooms along the Indian coasts (Padmakumar et al. 2018; Baliarsingh et al. 2018; Kumar et al. 2020), and the dominance of cysts (if any) might pose a concern and needs a vigilant approach as there is a rising trend in the incidences of HAB events in the Indian exclusive economic zone (Padmakumar et al. 2012). Cochin and New Mangalore port are highly at risk to future HABs event compared to Kandla port as most HAB species are dominant in this region (Fig. 3). Findings from Satta et al 2010, suggests Blixaea quinquecornis may be a benthic pelicle cyst as far as the surface sediment observation was concerned, and also recorded their apparently viable in deeper sediments (6-7cm). The planktonic form of B. quinquecornis have also been the reported in Cochin (Rodrigues et al. 2019) and New Mangalore port from the water coloumn analysis. Altough *B. quinquecornis* were reported in both Cochin and New Mangalore ports in the water column only in surface sediment of the latter was recorded. In this study *B. quinquecornis*, cyst shape observed was similar to that of its vegetative stages. This benthic pelicle cyst stages formed by *B. quinquecornis* need further investigation to study their life cycle. Studies indicated that the dinocysts act as a seed population, which in the dinoflagellates population dynamics play an essential role (Ishikawa and Taniguchi 2000). However, a detailed investigation needs to be commenced to monitor the dinocysts for predicting and preventing HABs events from other port (minor) along the coast of India.

Dinoflagellate cyst assemblages from the perspective of water quality

Several studies reported that dinocysts taxa could be used as indicators of water quality change and anthropogenic impact. To assess the water quality including eutrophication and pollution, indices such as species assemblage composition, species diversity, and species-richness are widely used (Hendey 1977; Wu 1984; Sommer 1995; Tsirtsis and Karydis 1998; Pospelova et al. 2002; Price et al. 2017; 2018). Shannon-Wiener diversity index has been used for pollution index in diatom communities (Hendey 1977), and dinocysts based Fisher's α and the number of species have been used as indicators of environmental conditions in estuarine systems (Pospelova et al. 2002, 2005). In the present study, the highest values for community indices were found to Cochin, followed by New Mangalore and Kandla port (Fig. 5). The low values in the Kandla port could be attributed to low planktonic biomass, high suspended load, high tidal amplitude, and sandy sediment texture in the region (Table 1). The tropical waters, characterized by low abundance and highly diverse dinoflagellate populations, generally expect high diversity and many species. However, the total number of species recorded is comparable between Cochin and New Mangalore ports and the other geographical regions except for Kandla, where the lowest species were recorded (Table. 4). The comparison of diversity and number of species values with available reports from temperate and tropical regions, including earlier studies from the same region, shows that the value of the present study is much lower than that found in the other areas (Table. 4). The decrease in diversity and number of species of dinocyst taxa can be used as a general indicator of eutrophication and pollution (Pospelova et al., 2002, 2005). Also, the decline in Fisher's a index and the number of species of dinocysts can also be considered a combined signal of toxic or organic pollution in the coastal regions (Pospelova et al. 2005). Therefore the low values found here could be attributed to the moderate to a high state of eutrophication and pollution in the port region. The values in the present study are comparable to the ranges reported from the coast of the Philippines (Baula et al. 2011, Table 4), and thereby the studied ports. According to the scaling index, as proposed by Pospelova et al. (2002), and Hendey (1977), Cochin, New Mangalore, and Kandla ports can be categorized as moderately to the highly polluted environment depending on the dinocysts diversity indices. Fisher's α and the number of species values obtained in Cochin port, New Mangalore, and Kandla port are lower than the values stated by Pospelova et al. (2002, 2005) indicated that the ports are polluted. Further, the scaling done with these proxies is on par with TRIX scores (for Cochin port - Rajaneesh et al. 2015 and New Mangalore port - Rath et al. 2019), pollution load index (for Cochin port; Chakraborty et al. 2014), and metal concentrations (for Cochin port; Martin et al. 2012). From the observation made, it is apparent that the diversity indices of dinocysts form useful environmental proxies.

Additionally, dinocyst taxa composition responses is different for eutrophicated systems (Price et al., 2017). For instance, in regions like Sishili Bay, Yellow Sea-China (Liu et al. 2012), British Columbia-Canada (Radi et al. 2007, Krepakevich and Pospelova 2010), Northeast Pacific margin (Radi and de Vernal 2004), Tokyo Bay-Japan (Matsuoka et al. 2003), and Northern Gulf of Mexico, (Price et al. 2018), the heterotrophic dominance of dinocyst taxa like Archaeperidinium minutum, Brigantedinium spp., Polykrikos kofoidii, and Protoperidinium leonis are known to reflect nutrient availability and changes in eutrophication. Usually, in coastal waters, heterotrophic dinoflagellates increase has referred to indicators for eutrophic environments (Dale et al., 1999; Matsuoka et al. 1999; Dale 2009; Price et al. 2018). In our study, more cysts were observed at medium and high eutrophication levels. Still, instead of heterotrophic autotrophic forms were the dominant (Fig. 5a) suggesting that former need not always be dominant in the eutrophic system, as observed in Cochin and New Mangalore ports. The dominance of autotrophic forms observed for most of the year in Cochin and New Mangalore ports were in contrast with other studied regions in the northern sector along the Indian coast, i.e., Mumbai port (northwest coast), Goa port (central west coast), and Visakhapatnam port (Northeast coast) (D'Costa et al., 2008; D'Silva et al., 2012). This dominance of autotrophic cysts does signify its higher planktonic production (Dale, 1983; Anderson et al., 1995; Joyce et al., 2005) and are reflected in this study. The planktonic dinoflagellate data for Cochin port (Rodrigues et al. 2019) and New Mangalore port (Rath et al. 2018) also showed the prevalence of autotrophs than heterotrophs. Such dominance is because of the changes driven by monsoon events (eg. nutrient-rich freshwater influx, the influence of upwelled water; Verlecar et al. 2006; Harnstrom et al. 2009; Madhu et al., 2010a and 2010b), which fuels phytoplankton production compared to the other studied regions on the northern side. While in the Kandla port, the dominance of the heterotrophic form was observed as they can tolerate broader salinity stress (Shin et al. 2010, 2011). Thus, in Kandla port, the high suspended load, along with the low chlorophyll levels, could be attributed to heterotrophic dominance (Table. 1). Further, the presence of the relatively high abundance of indicator taxa for eutrophic such as Pentaspharsodinium dalei, Lingulodinium polyedra, Polykrikos kofoidii/schwartzii, and the number of Protoperidinium species do emphasize that the Cochin, New Mangalore, and Kandla port are eutrophic (Dale et al., 1999; Pospelova et al. 2005; Radi et al. 2007; D'Silva et al. 2013; Price et al. 2018). Furthermore some of the dinocyst (such as Bitectatodinium spongium, Gonvaulax digitalis, Gymnodinium microreticulatum, Polykrikos kofoidii, P. schwartzii, and Protoperidinium stellatum) were relatively morphologically smaller in this study than available reports from other regions (Table 1) suggesting further investigation to delineating influencing factors. Nevertheless, the dominance of the dinocyst (Protoperidinium leonis, Protoperidinium conicum, Protoperidinium pentagonum, Gonyaulax spinifera complex. Lingulodinium polyedra, and Pyrophacus steinii) in the sediments of the complex port ecosystem will add information to a growing body of knowledge on dinoflagellate distribution patterns (Marret and Zonneveld 2003; Zonneveld et al. 2013) but also as an indicator of water quality.

Conclusion

This study provides an overview of dinoflagellate cyst assemblage (including new records and HAB species) with respect to trophic index (TRIX) for eutrophication from the major ports located on the west coast of India. The findings suggested that the dinoflagellate cyst abundance and species number showed a decreasing trend with respect to the geographical location of the ports (i.e., from south to north). Interestingly, the cyst abundance, species number, and the number of HAB species were highest in highly eutrophicated (Cochin) port followed by medium (New Mangalore) and low (Kandla) eutrophicated ports. The dominance of autotrophs in eutrophicated Cochin and New Mangalore port confirms that heterotrophs need not always be dominant and this is in contrast to offshore regions where heterotrophs such as Protoperidinium tend to be dominant. However, more data points are required to ascertain heterotrophs dominance in the offshore location along the Indian Furthermore, the relatively high indicator cyst taxa (Lingulodinium, Polykrikos, and coasts. Protoperidnium spp) confirm a eutrophic system. Additionally, the investigation records six new species, including a potential HAB species in the dinocyst checklist from the region. This study demonstrates the utility of dinoflagellate cyst assemblage (species numbers, fisher's a index, and indicator species) as potential indices for eutrophication in monsoon-influenced ports systems. Cyst assemblages also emphasize greater HAB risks in the high trophic index ports (Cochin and Mangalore) than the low trophic index port (Kandla) under favorable conditions.

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Table 1. Ranges of environmental parameters recorded from Cochin, New Mangalore, and Kandla ports. * The TRIX score during the sampling period, which conducted continuously over 3-4 days encompassing different tidal regimes and associated changes, didn't vary significantly for either of the three ports and this strengthen the utility of TRIX index.

		Ports	
Parameters	Kandla	New Mangalore	Cochin
Temperature (°C)	26.5 ± 3.6	27.9 ± 3.6	28.7 ± 1.7
Salinity (psu)	36 ± 6	35.2 ± 0.8	20.8 ± 11.3
Dissolved Oxygen (mg/L)	5.7 ± 0.8	3.4 ± 1.8	4.6 ± 2
Phosphate (µM)	4 ± 1.2	2.6 ± 1.3	3.3 ± 1.3
Silicate (µM)	72.7 ± 37.3	22.1 ± 14.2	37.7 ±16.7
Ammonia (µM)	5.7 ± 5.9	28.3 ± 10	28.2 ± 12.9
Nitrite (µM)	5.4 ± 2.1	2.8 ± 2	0.9 ± 0.7
Nitrate (µM)	12.6 ± 5.6	4.7 ± 4.2	4.7 ± 3.1
Chl a (µg/L)	0.6 ± 0.3	8.8 ± 9.3	20.6 ± 22.2
TRIX score*	$3.5\pm\ 0.7$	4.6 ± 0.9	5.5 ± 0.8
Depth (m)	9-10	7.5-14	9.75-13.7
Sediment Texture (%)	Sandy (55- 70%)	Silty- clayey (58-64%)	Silty- clayey (55-67%)

Table 2. List of occurrence of potentially harmful dinoflagellates from three ports with brief description.

Sr. No	Species name	Effect of bloom	Toxin type	Cochin	Mangalore	Kandla
	Autotrophs					
1	Alexandrium tamarense complex		Saxitoxin	+	+	
2	Cochlodinium sp		Possible Ichthyotoxic	+	+	
3	Gonyaulax scrippsae	Discolouration of water and oxygen depletion		+	+	
4	Gonyaulax spinifera		YTX	+	+	+
5	Lingulodinium polyedra		YTX	+	+	+
6	Protoceratium reticulatum		YTX	+	+	+
7	Scrippsiella acuminata	Discolouration of water		+	+	
8	Pyrodinium bahamense		Saxitoxin	+		
	Heterotrophs					
9	Blixaea quinquecornis	Discolouration of water and oxygen depletion			+	

Table 3. Checklist of dinoflagellates cyst types for six major ports located along the west coast of India. Values indicate maximum abundance (cysts g^{-1} dry sediment) recorded. The initial d, l, b, and p indicates the diameter, length, breadth, and process length of the dinocyst. Bold species names indicate smaller sized dinocyst.

Species list Mumbai Goa Cochin Kandla Mangalore Cochin Joint (and) Joint (and) <thjoint (and)<="" th=""> <thjoint (and)<="" th=""></thjoint></thjoint>		D'sil	va et al 2	011		Present stud	y	Present study Size (um)	Literature Size (um)
Autotrophs Image: second symplex index in the symplex index in the symplex index inde	Species list	Mumbai	Goa	Cochin	Kandla	Mangalore	Cochin	Size (µiii)	
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$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	affiine*	2	2	57					
minutum * 2 8 80 \sim \sim \sim Alexandrium tumarense 3 37 19 78 $d=45.47$ Complex* 3 37 19 78 $d=45.47$ Alexandrium spp 27 $d=40.50$, spongium $d=40.50$, $p=8.14$ $p=10^{-1}$ Cochlodinium cf. polykrikoides* 9 $d=40.50$ $d=40.50$ $d=60.73$ Gonyaulax digitalis 11 74 2 83 37 $p=10^{-1}$ Gonyaulax digitalis 11 1 74 2 83 37 $p=10.2^{-1}$ $p=50^{-1}$ Gonyaulax digitalis 11 1 74 2 83 37 $p=10.12$ $p=5.9^{-1}$ Gonyaulax scrippsæ* 45 29 27 42 271 $p=10.12$ $p=5.9^{-1}$ Gonyaulax sprinjera complex * 72 28 148 2 820 2561 $p=12.17^{-1}$ Gonyaulax spp 10 107 36 79	Alexandrium								
Alexandrium tumarense complex* 3 37 19 78 Alexandrium spp 27 6 6 Bitectatodinium spp 1 9 78 Bitectatodinium spp 1 9 6 6 Cochlodinium cf. polykrikoides* 9 9 1 9 6 Cochlodinium sp 10 19 49 d=40-50 6 6 Gonyaulax 1 1 19 49 d=40-50 1 6 6 5 6 9 1 1 1 1 1 1 9 9 1	minutum *	2	8	80					
tamarense complex* 3 37 19 78 Alexandrium spp 27 <td< td=""><td>Alexandrium</td><td></td><td></td><td></td><td></td><td></td><td></td><td>d= 45-47</td><td></td></td<>	Alexandrium							d= 45-47	
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	tamarense								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	complex*	3		37		19	78		
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Alexandrium spp			27					
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Bitectatodinium							d= 40-50,	d= 63
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	spongium				1			p= 8- 14	p= 10 ⁻¹
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Cochlodinium cf.								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	polykrikoides*		9						
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Cochlodinium sp		10			19	49	d= 40-50	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $								d= 30,	l=60-73
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Gonyaulax							p= 10	b=45-63
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	digitalis	11	1	74	2	83	37		p=15-25 ¹
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $								l= 40-45,	1 = 40-59
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Gonyaulax						10	b= 30-35,	b=26-42
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	elongatum					1	49	p=10-12	$p=5-9^{-1}$
Actropycle* 4.3 29 21 42 211 $p = 10-12$ Gonyaulax spinifera complex 72 28 148 2 820 2561 $d = 35-40$, $p = 13-15$ $p = 34-57$ * 72 28 148 2 820 2561 $p = 13-15$ $b = 34-57$ Gonyaulax spp 10 107 36 79 $p = 12-17^{-1}$ Gonyaulax spp 10 107 36 79 $p = 12-17^{-1}$ Gonyaulax spp 10 107 36 79 $p = 12-17^{-1}$ Gonyaulax spp 10 107 36 79 $p = 12-17^{-1}$ Gonyaulax spp 10 107 36 79 $p = 12-17^{-1}$ Gonyaulax sopinium microreticulatum 2 $p = 10^{-1}$ $p = 12-17^{-1}$ $p = 12-17^{-1}$ Gymnodinium impudicum 8 16 45 $d = 30-35$ $d = 36-62^{-1}$ Lingulodinium polyedrum * 22 10 44 1 233 565 $p = 10-12$ $p = 2-21^{-1}$ Lingulodinium sp2 8 9 4	Gonyaulax	45	20	27		42	271	d = 30-35,	
Gonyalitation $1=34-30$ $1=34-30$ $p=13-15$ $b=34-57$ $gpinifera$ complex 72 28 148 2 820 2561 $p=12-17^{-1}$ Gonyaulax spp 10 107 36 79 $p=12-17^{-1}$ Gymnodinium cf. 2 10 107 36 79 $p=12-17^{-1}$ Gymnodinium filter 2 10 107 36 79 $p=12-17^{-1}$ Gymnodinium filter 2 10 107 36 79 $p=12-17^{-1}$ Gymnodinium filter 2 10^{-1} 10^{-1} 10^{-1} 10^{-1} 10^{-1} Gymnodinium filter 1^{-1} 1^{-1} 10^{-1} 10^{-1} 10^{-1} 14^{-1} 233^{-1} 10^{-1}	scrippsde *	45	29	27		42	271	p=10-12	1-24.50
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	spinifara complex							u = 33-40, n = 13 - 15	1-34-30 b-34-57
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	*	72	28	148	2	820	2561	p= 15- 15	$n = 12 \cdot 17^{-1}$
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Comunitarian	12	10	107		26	70		p= 12 17
$ \begin{array}{c ccccc} Cymnodinium (j) & 2 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1$	Gonyaulax spp		10	107	-	30	/9		
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	catenatum	2							
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Gymnodinium	2							
Gymnodinium impudicum 8 16 45 $d=30-35$ $d=36-62^{-1}$ Lingulodinium polyedrum * 22 10 44 1 233 565 $p=10-12$ $p=2-21^{-1}$ Lingulodinium sp2 8 27 $p=8-10$ $d=40-50$ $p=30-35$ $d=19-36$ Pentapharsodinium dalei 8 9 40 1 7 22 $p=5$ $p=1-8^{-1}$ Pheopolykrikos 6 176 $p=10$ $p=7-12^{-1}$ $d=30-45$ Protoceratium 4 8 6 176 $p=10$ $p=7-12^{-1}$ Pyrodinium $d=35-40$ $p=5-7$ $d=35-40$ $p=5-7$ $p=5-7$ Pyrodinium $d=35-40$ $p=5-7$ $p=5-7$ $p=5-7$ $p=5-7$ Pyrodinium $d=35-40$ $p=5-7$ $p=5-7$ $p=5-7$ $p=5-7$	microreticulatum				1				
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Gymnodinium							d= 30-35	d= 36-62 ¹
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	impudicum		8			16	45		
polyedrum *2210441233565 $p=10-12$ $p=2-21^{-1}$ Lingulodinium sp2827 $p=8-10$ $d=40-50$ Pentapharsodinium dalei89401722 $p=5$ $p=1-8^{-1}$ Pheopolykrikos hartmannii486176 $p=10$ $p=7-12^{-1}$ Protoceratium reticulatum *251120481443 $p=10$ $p=7-14^{-1}$ Pyrodinium bahamense4907700	Lingulodinium							d= 48-50	d= 31-54
Lingulodinium sp2 $d = 40-50$ $p = 8-10$ Pentapharsodinium dalei $d = 30.35$ $d = 30.35$ $d = 19-36$ $p = 5$ Pheopolykrikos hartmannii $d = 30.75$ $d = 30.75$ $d = 19-36$ $p = 5$ Pheopolykrikos hartmannii $d = 70.75$ $d = 50-60$ $d = 70.75$ $p = 7.12^{1}$ Protoceratium reticulatum * 25 1 120 120 4 81 6 413 Pyrodinium bahamense var compressum* $d = 40.45$ $d = 35-40$ $p = 5-7$ $d = 35-40$ $p = 5-7$	polyedrum *	22	10	44	1	233	565	p= 10-12	p= 2-21 ¹
Lingulodinium sp2 8 27 $p=8-10$ Pentapharsodinium d= 30-35 d= 19-36 dalei 8 9 40 1 7 22 $p=5$ $p=1-8^{-1}$ Pheopolykrikos d= 70-75 d= 50-60 $p=7-12^{-1}$ $d= 40-45$ d= 38-48 Protoceratium 6 176 $p=10$ $p=7-12^{-1}$ Protoceratium $d= 40-45$ $d= 38-48$ $p=7-14^{-1}$ Pyrodinium $d= 35-40$ $p=5-7$ bahamense var $d= 35-40$ Pyrodinium 7								d= 40-50	
Pentapharsodinium d= 30-35 d= 19-36 dalei 8 9 40 1 7 22 $p=5$ $p=1-8^{-1}$ Pheopolykrikos 6 176 $p=10$ $p=7-12^{-1}$ Protoceratium 6 176 $p=10$ $p=7-12^{-1}$ Protoceratium $d=40-45$ $d=38-48$ reticulatum * 25 1 120 4 81 443 $p=10$ $p=7-14^{-1}$ Pyrodinium $d=35-40$ $p=5-7$ Pyrodinium 7 Pyrodinium 7 Pyrodinium 7	Lingulodinium sp2					8	27	p= 8- 10	
dalei 8 9 40 1 7 22 $p=5$ $p=1-8^{-1}$ Pheopolykrikos d=70-75 d=50-60 $p=7-12^{-1}$ d=70-75 d=50-60 $p=7-12^{-1}$ Protoceratium d=40-45 d=38-48 $p=10$ $p=7-14^{-1}$ Pyrodinium d=35-40 $p=5-7$ $p=5-7$ $p=5-7$ Pyrodinium $p=7-14^{-1}$ $p=5-7$ $p=5-7$ $p=5-7$ Pheopolysinium $p=7-14^{-1}$ $p=7-14^{-1}$ $p=5-7$ $p=5-7$ Pyrodinium $p=7-14^{-1}$ $p=7-14^{-1}$ $p=5-7$ $p=5-7$ $p=5-7$ Pyrodinium $p=7-14^{-1}$ $p=7-14^{-1}$ $p=7-14^{-1}$ $p=7-14^{-1}$ $p=7-14^{-1}$ $p=7-14^{-1}$	Pentapharsodinium	-				_		d= 30-35	d= 19-36
Pheopolykrikos d= 70-75 d= 50-60 hartmannii 4 8 6 176 p= 10 p= 7-12 1 Protoceratium d= 40-45 d= 38-48 p= 10 p= 7-14 1 Pyrodinium d= 35-40 p= 5-7 d= 50-60 Pyrodinium pyrodinium d= 35-40 p= 5-7 Pyrodinium 7 7 p= 10 p= 10	dalei	8	9	40	1	1	22	p= 5	$p=1-8^{-1}$
narmannu 4 8 6 176 $p=10$ $p=7-12^{-1}$ Protoceratium 25 1 120 4 81 $d=40-45$ $d=38-48$ pyrodinium d= 35-40 p= 7-14^{-1} d= 35-40 p= 5-7 d= 5-7 Pyrodinium pyrodinium 7 7 7 7	Pheopolykrikos	4	0			6	176	d = 70-75	d = 50-60
Protoceratium $d = 40-45$ $d = 38-48$ reticulatum * 25 1 120 4 81 443 $p = 10$ $p = 7-14^{-1}$ Pyrodinium $d = 35-40$ $p = 5-7$ $p = 5-7$ $p = 5-7$ $p = 5-7$ Pyrodinium $p = 10$ $p = 5-7$ $p = 5-7$ $p = 5-7$ $p = 5-7$ Pyrodinium $p = 5-7$	hartmannii	4	8			6	1/6	p=10	$p = 7 - 12^{-1}$
Perculation251120461443 $p=10$ $p=714$ Pyrodinium bahamense $d=35-40$ $p=5-7$ $p=5-7$ $p=5-7$	Protoceratium	25	1	120	4	Q 1	113	d = 40-45	d = 38-48 $p = 7.14^{-1}$
bahamense var p= 5-7 compressum* 4 Pyrodinium 7	Pyrodinium	23		120	4	01	440	p = 10 d = 35-40	P- /-14
compressum* 4 Purchase Purchase 7	hahamense var							n = 5-7	
Pyrodinium bahamense 7	compressum*	4						P^{-3-7}	
bahamense 7	Pyrodinium	-	1						
	bahamense						7		

Contd..

Species list	Mumbai	Goa	Cochin	Kandla	Mangalore	Cochin	PS (µm)	PR (µm)
Autotrophs								
Autotrophs							d= 65-95	1=80-110
							p = 10-15	b= 50-55
Pyrophacus steinii	29	17	18		919	1230	-	p= 8-13 ¹
							d= 85	
Pyrophacus sp						28	p= 10-15	
Scrippsiella trifida		16						
Scrippsiella							l= 38-40	
acuminata*	19	64	28		17	65	b=32-35	
Scrippsiella spp.		1			4	208	d= 35	
Hotorotrophs								
neterotropiis							1= 35-40	
Blixaea							b = 30-35	
quinquecornis *					17		p= 2-5	
Brigantedinium spp.				53			d= 38-40	d= 29-53 ¹
Dinlonelta name		10						
Diplopelta parva		12			+			
*	2							
T 1 1 1 1	24		50					
Lebouraia minuta	34		50				1-68.70	1 - 72 125
							h = 38-40	h = 48-63
Polykrikos kofoidii	7	16	74		2	118	p=5-10	$p = 6 \cdot 11^{-1}$
							l= 55-60	l = 60-72
							b=50	b= 44-55
Polykrikos schwartzii	4	10	148		22	209	p= 5-7	p= 8.5-14 ¹
Polykrikos sp		9						
Protoperidinium							d= 40-45	
avellana				1				
							l= 35-38	
Protoperidinium	4	14	20		25	220	b=30-32	
Protoperidinium	4	14	50		2.5	229	p = 3 - 3 1 - 85 - 90	
compressum	8	9	43		38	107	b=70-75	
Protoperidinium							d= 40-45	d= 29-49 ¹
conicoides	10			1				
							l= 70-75	
Protoperidinium	20	22	02	2	22	279	b = 75-80	
Protoperidinium	39		02	5		278	p=10-12 d=40-45	
denticulatum		8		1			u- 40 45	
							l= 70-80	
Protoperidinium							b= 60-70	
divaricatum	4	9			1	14	p= 10-15	
Protoperidinium	15	4.4	95	1	196	409	l = 80-90	
Protoporidinium	15	44	85	1	186	498	b = 75-85 1 = 60.70	1-60.801
leonis	36	47	241	4	371	1115	h = 45-55	1-00-00
Protoperidinium		.,			0,1		l= 70-75	
oblongum	49	45	108		154	397	b= 65-70	
Protoperidinium							l= 70-95	
pentagonum	59		28		344	717	b= 65-85	
Protoperidinium cf.	27	40	102					
pentagonum	5/	48	103					

Contd..

Species list	Mumbai	Goa	Cochin	Kandla	Mangalore	Cochin	PS (µm)	PR (µm)
Heterotrophs								
Protoperidinium							l= 60-65	d= 48-60 ¹
subinerme	10	37	60	1	6	21	b= 50-55	
							l= 55-60	
Protoperidinium sp.1	4	8	2			11	b= 50-55	
Protoperidinium sp.2	2							
Protoperidinium sp.3								
Protoperidinium sp.4		4						
Protoperidinium sp.5		21	37					
Protoperidinium							l= 85-90	l = 92-125
stellatum					40	72	b= 85-90	b= 82-126 ¹
Protoperidinium spp.		53	80					
Preperidinium							d= 55-60	
meunieri	6	51	111	0	103	119	b=48-50	

¹ https://www.marum.de/en/Karin-Zonneveld/Modern-Dinocyst-Key.html

Table 4. Comparison of dinoflagellate cyst species richness and diversity along the Indian coast and other regions.

	No.	Dive	ersity	Grandar	Dellasticas have d	
Region	species	Shannon- Weiner	Fisher's index	richness on diversity		Reference
Kandla Port,						
West coast of India	15	0.6 - 1.3	0.2 - 2.6	1 - 4	moderate to high	Present study
New Mangalore Port, West coast of India	33	0.6 - 2.2	0.3 - 3	1 - 14	moderate to high	Present study
Cochin Port,						
West coast of India	35	0.5 - 2.6	0.3 - 2.8	2 - 18	moderate to high	Present study
Cochin Port,						D'Silva et al.,
West coast of India	29	1.6-2.5	-	-	low to moderate	2011
Mumbai Port,						D'Silva et al.,
West coast of India	32	1-2.6	-	-	low to moderate	2011
Goa Port,						D'Silva et al.,
West coast of India	35	0.5-2.5	-	-	moderate to high	2011
Vishakapatnam Port,						D'Silva et al.,
East coast of India	28	0.4-2.4	-	-	moderate to high	2012
Buzzards bay,						
Massachusetts,					low	Pospelova et al.,
East coast of USA	37	-	3.5-6.4	12 - 26		2005
Bolinao, Pangasinan,						Baula et al.,
West coast of Philippines	34	0.8-2.5	0.6-3.7	3 - 15	moderate to high	2011

Figure 1. Location of sampling stations in different ports. a) Kandla port, b) New Mangalore port, and c) Cochin port along the west coast of India. The details of the sampling stations are provided in the Supplementary Tables 1 to 3.

Figure 2 a. Micrographs of HAB dinoflagellates (1) Alexandrium tamarense complex, (2) Cochlodinium sp, (3) Gonyaulax scrippsae, (4) Gonyaulax spinifera complex, (5) Lingulodinium polyedrum, (6) Lingulodinium sp2, (7) Pyrodinium bahamense, (8) Scrippsiella acuminate, (9) Blixaea quinquecornis. Scale bar is 20um.

Figure 2 b. Micrographs of dinoflagellates (1) *Bitectatodinium spongium*,(2) *Gonyaulax digitalis*, (3) *Gonyaulax elongatum*, (4) *Gymnodinium microreticulatum*, (5) *Pentapharsodinium dalei*, (6) *Pheopolykrikos hartmannii*, (7) *Pyrophacus steinii*, (8) *Pyrophacus* sp1, (9) *Scrippsiella sp*, (10) *Brigantedinium* spp., (11) *Polykrikos kofoidii*, (12) *Polykrikos schwartzii*, (13) *Protoperidinium avellana*, (14) *Protoperidinium claudicans*, (15) *Protoperidinium compressum*, (16) *Protoperidinium conicoides*, (17) *Protoperidinium conicum*, (18) *Protoperidinium denticulatum*, (19) *Protoperidinium divaricatum*, (20) *Protoperidinium latissimum*, (21) *Protoperidinium leonis*, (22) *Protoperidinium oblongum*, (23) *Protoperidinium pentagonum*, (24) *Protoperidinium* sp1, (25) *Protoperidinium stellatum*, (26) *Protoperidinium subinerme*, (27) *Preperidinium meunieri*. Scale bar is 20um.

Figure 3. Dinoflagellate cysts abundance (cysts g^{-1} dry weight) in the three ports Kandla (K), New Mangalore (M) and Cochin (C). * indicates potential HAB species. Largest bubble corresponds to 4000 cysts g^{-1} dry weight

Figure 4.Variations in dinocyst (a) abundance, (b) autotrophic and heterotrophic forms based on nutrition (c) species-richness, (d) Shannon-Wiener's diversity, (e) Fisher's alpha (α) index and (f) Species number from the Kandla, Cochin and New Mangalore ports.

Figure 5. Relationship between trophic index (TRIX) with the total community and their nutrition mode forms (i.e. Autotrophs and Heterotrophs) (a) abundance, (b) species number, (c) species-richness, (d) Fishers index and (e) Shannon-Wiener's diversity of the three ports. For motile-stages and cysts the TRIX scores for surface and near bottom water respectively were used. LE, ME and HE refers to low, medium and high eutrophication, respectively.

Figure 6. Dendogram of dinoflagellates cysts from sediments with respect to (a) ports and (b) species. Species codes are as follows *Alexandrium tamarense* complex (Ale Tam), *Bitectatodinium spongium* (Bit spo), *Cochlodinium sp* (Coc sp), *Gonyaulax digitalis* (Gon dig), *Gonyaulax elongatum* (Gon elo), *Gonyaulax scrippsae* (Gon scr), *Gonyaulax spinifera* complex (Gon spi), *Gymnodinium microreticulatum* (Gym mic), *Lingulodinium polyedrum* (Lin pol) *Lingulodinium sp2* (Lin sp), *Pentapharsodinium dalei* (Pen dal), *Pheopolykrikos hartmannii* (Phe har) *Protoceratium reticulatum* (Prot rec), *Pyrodinium bahamense* (Pyr bah) *Pyrophacus steinii*, (Pyr ste), *Pyrophacus* sp1 (Pyr sp), *Scrippsiella acuminate* (Scr acu), *Scrippsiella sp* (Scr sp), *Brigantedinium avellana* (Pro ave), *Protoperidinium claudicans* (Pro cla), *Protoperidinium compressum* (Pro com), *Protoperidinium conicoides* (Pro con) *Protoperidinium conicum* (Pro con), *Protoperidinium divaricatum* (Pro div), *Protoperidinium latissimum* (Pro lat), *Protoperidinium sp1* (Pro sp1), , *Protoperidinium stellatum*, (Pro ste), *Protoperidinium subinerme* (Pro sub), *Preperidinium meunieri* (Pre meu).



Fig. 1.



Fig. 2a.



Heterotrophs





Fig 3.



Fig 4.



Fig 5.



Fig.6

Stn.	Stn. Name	Latitude	Longitude
No.		(°N)	(°E)
1	Custom Buoy	9.968	76.253
2	Container Terminal	9.975	76.252
3	Fishery Harbour	9.940	76.263
4	L-Jetty/Dry Dock	9.945	76.267
5	South coal berth	9.953	76.267
6	Qua-1	9.954	76.267
7	Qua-4	9.958	76.265
8	North Coal Berth	9.964	76.261
9	Boat Train Pier	9.965	76.260
10	DC Jetty	9.969	76.264
11	Qua-6	9.967	76.267
12	Qua-8	9.966	76.270
13	Qua-10	9.964	76.275
14	Ro-Ro Jetty	9.960	76.278
15	Naval Jetty	9.957	76.281
16	Cochin Shipyard	9.955	76.286
17	Bunker Oil Jetty	9.958	76.285
18	IFP Jetty	9.960	76.284
19	South Tanker Berth	9.962	76.280
20	North Tanker Berth	9.964	76.279
	Ernakulam Ferry		
21	Jetty	9.971	76.279
22	Cochin Oil Terminal	9.970	76.270
23	Ernakulam Creek	9.978	76.275

Suppl. Table no.1. Details of sampling stations in Cochin harbour.

Stn.	Stn. Name	Latitude	Longitude
No.		(°N)	(°E)
1	General cargo berth	12.929	74.820
2	General cargo-1	12.930	74.819
3	General cargo-2	12.932	74.819
4	General cargo-3	12.934	74.818
5	Northern return	12.934	74.818
6	General cargo-4	12.933	74.817
7	General cargo-5	12.932	74.817
8	General cargo-6	12.930	74.820
9	Iron ore berth	12.925	74.819
10	POL/Liquefied Gas berth	12.922	74.811
11	Crude/POL berth-1	12.919	74.813
12	Crude/POL berth-2	12.916	74.813
13	Crude/POL berth-3	12.917	74.815
14	Under construction	12.920	74.815
15	Multipurpose berth	12.928	74.816
16	Coal berth	12.927	74.812
17	Channel marker buoy-1	12.926	74.808
18	Channel marker buoy-2	12.924	74.808
19	Turning circle	12.925	74.814

Suppl. Table no.2. Details of sampling stations in New Mangalore port.

Stn. No.	Stn. Name	Latitude	Longitude
	IOC Jetty	23°02'31.3"N	70°13'20.0"E
2	IFFCO Jetty	23°02'17.0"N	70°13'20.8"E
3	Rajiv Jetty	23°02'04.1"N	70°13'24.3"E
4	Indira Jetty	23°01'54.4"N	70°13'22.9"E
5	Shastri Jetty	23°02'12.6"N	70°13'23.0"E
6	Nehru Jetty	23°02'00.9"N	70°13'23.2"E
7	Local Fishing craft Anchorage-1	23°01'16.3"N	70°13'20.5"E
8	Local Fishing Craft Anchorage-2	23°01'03.0"N	70°13'19.0"E
9	Dry Dock	23°00'45.10"N	70°13'21.6"E
10	Crafts Berthing Jetty	23°00'26.0"N	70°13'24.6"E
11	Cargo Berth-1	23°00'19.7"N	70°13'25.7"E
12	Cargo Berth-3	23°00'13.3"N	70°13'26.4"E
13	Cargo Berth-5	23°00'01.8"N	70°13'26.0"E
14	Cargo Berth-7	22°59'44.7"N	70°13'27.1"E
15	Cargo Berth-9	22°59'32.0"N	70°13'28.3"E
16	Cargo Berth-10	22°59'29.8"N	70°13'28.8"E
17	Cargo Berth-11	22°59'23.7"N	70°13'30.2"E
18	Cargo Berth-12	22°59'17.3"N	70°13'29.5"E
19	Cargo Berth-14	22°59'02.4"N	70°13'35.5"E
20	Cargo Berth-15	22°58'53.3"N	70°13'39.5"E
21	Kandla Creek-1	22°58'56.7"N	70°14'06.6"E
22	Kandla Creek-2	22°59'32.5"N	70°13'55.5"E
23	Kandla Creek-3	23°0'53.28"N	70°13'54.39"E
24	Kandla Creek-4	23°02'10.2"N	70°13'50.5"E
25	Kandla Creek-5	23°2'33.80"N	70°13'35.10"E
26	Kandla Creek-6	23°02'52.0"N	70°13'30.9"E

Suppl. Table no.3. Details of sampling stations in Kandla port.