# Distinct environmental parameters influence the abundance of living benthic foraminifera morphogroups in the southeastern Arabian Sea

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# Abstract

The ambient environmental parameters have a great bearing on the morphology of living flora and fauna. In this study, we tested this hypothesis on one of the most dominant group of living unicellular marine microorganism, benthic foraminifera, in the dynamic region of the southeastern Arabian Sea. The living benthic foraminifera from 43 surface samples collected between 25 to 2980 metres of water depth were segregated into eight morphogroups (tapered/cylindrical, flattened-ovoid, biconvex, planoconvex, flattened-tapered, spherical, rounded-trochospiral, and rounded-planispiral). We report that the high organic carbon availability combined with deficiency of oxygen results in benthic foraminifera thriving in the oxygen-rich bottom waters have a high surface area to volume ratio, commonly reported as epifuanal, consisting of biconvex and planoconvex forms. Additionally, we also report that the abundance of other morpho-groups, namely flattened-tapered, spherical, rounded-trochospiral, and rounded-planispiral, is also controlled by the distinct environmental parameters. We suggest that the living benthic foraminifera are an excellent indicator of the ambient environmental parameters and can be used to reconstruct paleoenvironments.

Keywords: benthic foraminifera, morphogroups, ecology, Arabian Sea

#### Introduction

Everything in the cosmos has a distinct outward shape, which is referred to as morphology. All morphologies, from galaxies (Kelkar et al., 2017) to microscopic organisms (Chien et al., 2012), are shaped by its surrounding environmental parameters. The environmental parameters also control the volume and surface area of the organisms in an ecosystem and is often interpreted as its adaptability (Holt and Miller, 2010). A successful ecosystem is the result of effective adjustment of the biotic and abiotic components, where biotic component is represented by the living fauna and flora, and the abiotic components include the physico-chemical parameters. In case of marine ecosystem, benthos is one of the most successful living groups in the world ocean. The success of marine benthos in occupying diverse habitats is highly coupled with their adaptability in different kind of environmental conditions. The exoskeleton bearing unicellular microorganism benthic foraminifera are one of the most diverse and ubiquitous marine benthic organisms. Benthic foraminifera are found in a variety of marine environments, including brackish and sometimes freshwater also (Gooday, 2001; Saraswat and Nigam, 2013). The widespread distribution of benthic foraminifera is due to their remarkable adaptability to a variety of environmental conditions. Benthic foraminifera develop a variety of morphologies that are typically influenced by their ambient environmental parameters (Gooday, 2001; Prazeres et al., 2016; O'Brien et al., 2021). The distinct morphology representing a particular habitat as well as the excellent preservation of the exoskeleton, makes benthic foraminifera as one of the bestsuited and most frequently used groups to reconstruct the past environmental variations (Saraswat and Nigam, 2013). The abundance, distribution and morphology of benthic foraminifera in various oceanic settings is frequently linked to seawater environmental characteristics. Temperature, salinity, pH, water depth, dissolved oxygen, and food availability have all been identified as important environmental parameters affecting benthic foraminifera abundance and diversity (Jorissen et al., 1995; Singh et al., 2021; Suokhrie et al., 2021). Furthermore, benthic foraminifera have been shown to be one of the most prominent bio-indicators of marine pollution (Frontalini et al., 2011; 2015) due to their rapid response to ambient environmental changes.

Environmental parameters considerably modulate the morphology of benthic foraminifera. The species belonging to diverse genera have previously been clubbed to represent a distinct set of environmental parameters and referred as benthic foraminifera morphogroups (Chamney, 1976; Bernhard, 1986). Initially, benthic foraminifera were segregated into two broad morphogroups, namely epifaunal and infaunal benthic foraminifera based on their habitat (Corliss, 1985; 1991). Epifaunal benthic foraminifera reside close to sediment water interface, whereas infaunal benthic foraminifera were spotted burrowed up to a few centimetres deep in sediments (Jorissen et al., 1995; Singh et al.,

2018; Kaithwar et al., 2020). Epifaunal benthic foraminifera have high surface area to volume ratio and are usually abundant in oxygen rich waters on the shallow continental shelf as well as deep-water regions where bottom water replenishes the oxygen (De and Gupta, 2010; Khare et al., 2017). On the other hand, infaunal (occasionally also characterized as shallow infaunal, intermediate and deep infaunal) benthic foraminifera have low surface area to volume ratio and thrive well in low bottom water oxygen condition such as oxygen minimum zones (Nigam et al., 2007, Mazumder and Nigam, 2014). A few workers refer them as asymmetrical benthic foraminifera (AABF; sometimes also known as rectilinear forms; Nigam et al., 2007) and rounded symmetrical benthic foraminifera (RSBF) morphogroups (Manasa et al, 2016; Naik et al., 2017; Suokhrie et al., 2018). The benthic foraminifera belonging to AABF morphogroup are elongated, with the length exceeding the width of the specimen. The symmetrical forms, on the other hand, are usually rounded in shape. RSBF are abundant in the shallow waters with high energy conditions (Manasa et al., 2016) as well as in deep water where dissolved oxygen is abundant (Suokhrie et al., 2018). AABF, on the other hand, is primarily abundant in the intermediate water, where there is plenty of food but oxygen is limited (Singh et al., 2018; Suokhrie et al., 2021). Epifaunal-infaunal and AABF-RBSF, to a certain degree, represent same morphology based differentiation.

Later, it was realized that benthic foraminifera can be categorized further into several other morphgroups, namely, tapered/cylindrical, spherical, biconvex, planoconvex, flattened-ovoid, flattened-tapered, rounded-trochospiral, rounded-planispiral and milioline (Corliss and Chen, 1988; Corliss and Fois, 1990). The distribution of these morphogroups was correlated with water depth (Chamney, 1976; Severin, 1983; Corliss, 1985; Jones and Charnock, 1985; Bernhard, 1986; Corliss and Fois, 1990), as well as the organic carbon abundance (Corliss and Chen, 1988), microhabitat preferences (e.g., Corliss 1985; Corliss and Emerson, 1990). The relative abundance of elongateflattened, biconvex, and tapered forms increased with increasing water depth, but the planoconvex morphogroup's abundance decreased from shallow to deep water (Severin, 1983). The depth distribution of these nine benthic morphogroups from the Norwegian Sea was initially studied by Corliss and Chen (1988). The tapered-cylindrical morphogroup had a depth range of 0-2500 m, but was more abundant in the upper 1000 m. The flattened tapered shapes were erratic and only found in the upper 500 metres. The depth distribution of rounded-planispiral tests ranged from 20 to 3000 m, while the interval 500-1000 m had its highest abundance. The spherical morphogroup was less abundant, and was reported between 200-1600 m. The flattened-ovoid forms were found in a wide range of water depth (50-3200 m), with the highest abundance between 700 and 1500 m. The planoconvex morphogroup, although found between 0 and 4000 m, showed two peaks of abundance,

between 200 and 500 m, and in the interval of 1500-2500 m. The biconvex morphogroup was recorded at depths ranging from 0 to 4000 m, with the maximum abundance between 1500 and 4000 m. Milioline forms gradually increased from 100 to 3200 m, but as a whole they were less abundant. The rounded-trochospiral morphogroup ranged from 0 to 4000 m, but with low abundance throughout.

Such an elaborate distinction between different benthic foraminifera morphogroups has not been applied in the Indian Ocean, with a majority of the published works either using only two major categories (AABF and RSBF) (Nigam et al., 2007; Manasa et al., 2016; Suokhrie et al., 2018), or applying it to only the agglutinated benthic foraminifera (Saalim et al., 2019). However, recently Verma et al. (2018) categorized benthic foraminifera from a sediment core in to these nine morphogroups. It was reported that planoconvex and milioline morphogroups prefer oxygen rich environment, whereas, tapered/cylindrical forms were abundant during the period of strengthened oxygen minimum zone.

It is also worth noting that most of the earlier efforts at categorizing benthic foraminifera into different morphogroups, based on test morphology, used dead assemblages. The dead assemblages typically comprise of a mix of benthic foraminifera resultant from a long interval and most likely various microhabitats as well (Loubere and Gary, 1990). Furthermore, the selective breakdown because of dissolution and mechanical destruction significantly alters the original foraminiferal assemblage (Loubere, 1997). Therefore, the environmental preferences of the morphogroups should be assessed from the living benthic foraminifera assemblages. Additionally, the living benthic foraminifera (rose Bengal stained) are now widely practiced and accepted for bio-monitoring (FOBIMO-FOraminiferal BIo-Monitoring, Schönfeld et al., 2012; Frontalini & Coccioni, 2011; Saalim et al., 2017; Dubey et al., 2018; Jorissen et al., 2018). Therefore, here, we study the relationship between the different morphogroups of living benthic foraminifera and environmental parameters in a dynamic region of the southeastern Arabian Sea. The findings will be helpful in deciphering environmental preferences of the morphogroups in the present oceanic settings, which may be utilised to deduce past environmental conditions.

#### Physiography of the study area

The study region is in the northern Indian Ocean, where the Arabian Sea meets the Bay of Bengal. The seasonal monsoon winds in the Arabian Sea and low salinity water input from the Bay of Bengal are two oceanographic processes that have a significant impact on the biological as well as environmental parameters in the study region (Banse, 1959; Smitha et al., 2008). The area encompasses a large portion of the continental shelf as well as the slope region. Additionally, a relict carbonate platform (~60 and

200 m water depth) is found in the shelf area (Rao et al., 2003). The sediments are very coarse sand type in the shallow depths, and finer on the slope (Sivadas et al., 2020). The sea level shift in this region is particularly apparent as the area has a broad shelf in the central portion, then a sharp increase in depth, practically a vertical drop, followed by a continental slope. Aside from that, seasonal coastal currents bring warm, high-saline water to the study area during the southwest monsoon and cold, low-saline water during the northeast monsoon (Shankar et al., 2002).

#### **Material and Methodology**

The surface sediment samples (43 number) for living benthic foraminifera were collected during the post monsoon season in October 2014, onboard *Sindhu Sadhana* voyage SSD004 (Figure 1). The Ocean Scientific International Limited Maxi Multi-corer was used to collect the undisturbed core-top/surface samples along with overlaying water. The multi-corer tubes were 600 mm long with 110 mm outer diameter and 100 mm internal diameter. Additionally, the overlying water collected in the multi-core top was used to measure the seawater physico-chemical parameters including temperature, salinity and dissolved oxygen at the sediment-water interface. The sediments were sub-sampled at 1 cm interval. One half of each sub-sample in the top 10 cm, was immediately stained with rose-Bengal (2 g of rose-Bengal stain dissolved in 1 L of 70% ethanol). The stained samples were stored at 4°C for 7-8 weeks for proper staining and distinction between the living benthic foraminifera from the dead ones.



Figure 1: The surface sediment samples collected from the southeastern Arabian Sea. The coloured contours are bathymetry and elevation. The surface samples were collected along the transects T1-T4. MSL = Mean Sea Level.

The top two centimetre of the sediment samples were processed to segregate living benthic foraminifera. The samples were wet sieved by using 63 µm sieve to segregate benthic foraminifera from the sediment matrix. The coarse fraction was oven dried overnight at 45°C, weighed and stored in plastic vials. A representative aliquot of the coarse fraction was weighed and 300 benthic foraminifera were picked under Olympus SZX 12 stereo-zoom microscope. All the picked specimens were identified up to species level. Benthic foraminifera morphogroups were classified following Corliss and Fois (1990). Furthermore, canonical correspondence analysis (CCA) was computed by using MultiVariate Statistical Package software to find out the relationship between living benthic foraminifera morphogroups and the ambient environmental parameters.

For carbon and nitrogen analysis, ~5 g of sediment from the unstained half was freeze-dried and powdered by using a clean agate mortar pestle. The total inorganic carbon (TIC) in the sediment was analyzed by using coulometer (model CM 5015 CO2), and the total carbon and nitrogen was analyzed by using elemental analyzer (model FLASH 2000 Thermo Scientific). The organic carbon (%C<sub>org</sub>) was calculated by subtracting TIC from total carbon.

#### Results



#### Living benthic foraminifera

Figure 2: The absolute abundance of living benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The living benthic foraminifera were found in all the samples collected from different depths. The average living depth of benthic foraminifera was ~2 cm in the southeastern Arabian Sea (Kaithwar et

al., 2020). The absolute abundance of living benthic foraminifera in the top 2 cm varied between 3 and 448 specimen/g sediment (Figure 2). The highest abundance was on the slope of the Gulf of Mannar (T1 transect). The foraminiferal abundance was generally high at intermediate water depths along all transects. As compared to the intermediate depths, the shallow and deeper water sediments contained <50 specimen/g sediment. Thus the living benthic foraminiferal abundance at shallow and deeper depths was 3-4 times less than that at intermediate water depths. A total 174 living benthic foraminifera species belonging to eight morphogroups were found. The distribution pattern of each morphogroup is discussed below in detail.

#### **Tapered/Cylindrical**



Figure 3: The relative abundance of living tapered/cylindrical morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The specimens belonging to a total of 52 species (Table 1) were included in the tapered/cylindrical morphogroup. This morphogroup was the most common in the southeastern Arabian Sea. Its relative abundance ranged from being absent to 100.00%, with an average of 32.29%. The tapered and cylindrical forms were mostly abundant at the intermediate depths (1000-1500 m) of all transects. The highest relative abundance of the tapered and cylindrical forms was in the Gulf of Mannar transect (T1 transect). Furthermore, two deeper stations, MC-56 and 59, have a higher proportion of living tapered and cylindrical benthic foraminifera (Figure 3)

# Spherical



Figure 4: The relative abundance of living spherical morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The spherical morphogroup included 16 species in the southeastern Arabian Sea (Table 1). The abundance of spherical morphogroup ranged from being absent at several stations to a maximum of 14.72%, with an average of 3.26%. Interestingly, the spherical tests were relatively less abundant in the shallow water on the shelf and dominant at the deep water stations. Transect T3 had a higher percentage of living spherical forms, and shallow water stations G1 and MC10 also had a high abundance (Figure 4).

Biconvex



Figure 5: The relative abundance of living biconvex morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

Biconvex morphogroup includes 15 species (Table 1) and its relative abundance ranged from being absent at several stations to 40.00%, with an average of 8.91%. Generally, the abundance of biconvex forms increased on the upper continental slope in Transect T3 and T4. However, in Transect T1 and T2, the abundance was higher on the lower continental slope (Figure 5).

#### Planoconvex



# Figure 6: The relative abundance of living planoconvex morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

Planoconvex morphogroup comprised of specimens belonging to 15 species (Table 1). Its relative abundance ranged from being nearly absent to as high as 66.66%, with an average of 13.78%. Planoconvex forms were abundant in transects T2, T3 and T4, whereas less abundant in T1. The highest relative abundance of planoconvex morphogroup was found at station MC30 at a depth of 1506 m. Furthermore, the relative abundance was comparatively high at intermediate water depths in all transects (Figure 6).

## Flattened-ovoid



Figure 7: The relative abundance of living flattened-ovoid morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

Flattened-ovoid forms (13 species) were comparatively rare, with abundance ranging from being absent to 39.58%, with an average of 6.33%. It was abundant on the upper continental slope but comparatively scarce on the shelf and deeper slope regions. The highest abundance was at station MC27 at 764 m water depth (Figure 7).



# Flattened-tapered

Figure 8: The relative abundance of living flattened-tapered morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

Flattened-tapered morphogorup included 25 species (Table 1). It was moderately abundant in the southeastern Arabian Sea, with abundance ranging from nearly absent to 75.00%, with an average of 19.29%. The abundance of living flattened-tapered benthic foraminifera was comparatively higher on the continental shelf and deeper slope regions, whereas the intermediate water depth stations had a very few living flattened-tapered benthic foraminifera (Figure 8).



#### **Rounded-trochospiral**

Figure 9: The relative abundance of living rounded-trochospiral morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

In the southeastern Arabian Sea, rounded-trochospiral (17 species) relative abundance ranged from being absent to 42.85%, with an average of 8.93%. Barring a few exceptions, its relative abundance generally increased with depth in the transect T3 and T4. The highest abundance was at deep-water station MC55 (2238 m) (Figure 9).

## **Rounded-planispiral**



Figure 10: The relative abundance of living rounded-planispiral morphogroup of benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The overall abundance of rounded-planispiral morphogroup (18 species) was also moderate in the southeastern Arabian Sea. The abundance ranged from nearly absent to 33.33%, with an average of 7.03%. The abundance was higher on the continental shelf, upper slope, and intermediate slope, but low on the deeper slope. The highest abundance was at MC30 at a depth of 1506 m. In general, the relative abundance of living rounded planispiral forms increased with depth in the transect T4 (Figure 10).

# Infaunal



Figure 11: The relative abundance of living infaunal benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The species were grouped into infaunal and epifaunal forms, following Kaithwar et al (2020). The relative abundance of living infaunal benthic foraminifera was comparatively very high with the entire assemblage comprising of only infaunal forms at several stations. The average relative abundance of living infaunal benthic foraminifera was 68.21%. The relative abundance of infaunal forms was higher on the continental shelf and upper slope. Its abundance decreased rapidly after 1500 m in a few parts of the study area (Figure 11).

# Epifaunal



Figure 12: The relative abundance of living epifaunal benthic foraminifera in the southeastern Arabian Sea. The numbered contours are bathymetry.

The epifaunal benthic foraminifera were comparatively less abundant in the southeastern Arabian Sea. The abundance ranged from being absent at few stations to 71.42%, with a mean of 31.62%. The highest abundance was at MC55 (2460 m). The abundance was relatively high on the continental shelf and deeper slope, but was low at the intermediate depths (Figure 12).

#### **Environmental Parameters**



Figure 13: The temperature, salinity and dissolved oxygen in the water at the sediment-water interface in the southeastern Arabian Sea. The organic carbon concentration ( $%C_{org}$ ) and organic carbon to nitrogen ratio ( $%C_{org}/N$ ) in the top 2 cm of the sediments at all the stations is also plotted with depth (Modified after Singh et al., 2021). A) represents whole area, T1-T4) represents transect wise distribution of the environmental parameters.

Bottom water salinity, temperature, dissolved oxygen, organic carbon concentration (% $C_{org}$ ), organic carbon to nitrogen ratio (% $C_{org}$ /N) and water depth were the environmental parameters considered here to understand its effect on living benthic foraminifera morphogroups. The bottom water salinity varied

within a narrow range (34.7-35.1 psu) and decreased with depth. Bottom water temperature also decreased with water depth and ranged between 2.5 to 27.4 °C. The depth-wise trend of both the salinity and temperature was similar in all transects, although the range was different. A large transect to transect variation was observed in  $%C_{org}$ , although the samples collected from the intermediate depths in all transects had a higher organic carbon content. Organic carbon concentration in the sediment varied between 0.71 to 8.32%. The degradation of organic matter causes formation of perennial oxygen deficient zone and the value ranged between 0.35 to 4.25 mL/L. Additionally,  $%C_{org}/N$  value ranged between 4.8 to 17.8 (Singh et al., 2021). The  $%C_{org}/N$  varied the maximum transect to transect variation (Figure 13).





# Figure 14: The canonical correspondence analysis between different living benthic foraminifera morphogroups and ambient environmental parameters in the southeastern Arabian Sea.

The Canonical Correspondence analysis helped in finding out the correlation between morphogroups and ambient environmental parameters, where the direction and length of the arrow delineate positive/negative relationship. From the CCA plot, it was evident that the flattened-tapered form was positively correlated with bottom water salinity and temperature. This form was also negatively correlated with %C<sub>org</sub>/N and water depth. Similarly, epifaunal, biconvex and planoconvex forms were positively correlated with bottom water dissolved oxygen. These forms were also showing negative correlation with organic carbon percentage in the sediments. The spherical, rounded-planispiral and rounded-trochospiral forms were positively correlated with %C<sub>org</sub>/N and water depth, and negatively correlated with bottom water salinity and temperature. The infaunal, flattened-ovoid and tapered/cylindrical forms were positively correlated with %C<sub>org</sub> in the sediments. These forms were also negatively correlated with bottom water dissolved oxygen (Figure 14).

#### Discussion

From the surface distribution and CCA it was evident that several living benthic foraminifera morphogroups in the southeastern Arabian Sea were strongly modulated by the ambient environmental parameters. The major environmental parameters in the southeastern Arabian Sea included high primary productivity and concurrent perennial oxygen deficient zone formation, as well as the bottom water salinity, temperature, water depth, and  $%C_{org}/N$ .

A majority of the living benthic foraminifera in the study region were tapered and cylindrical and showed a positive correlation with %Corg or, in other words, food availability. The maximum abundance of such forms was between 1000-1500 m water depth, in contrast to other parts of the world ocean, such as the Norwegian Sea (Corliss and Chen, 1988) and the Gulf of Mexico (Corliss and Fois, 1990), where maximum abundance was found in the upper 1000 m. The difference in the depth zone of increased abundance of tapered and cylindrical foraminifera could be because the lower continental slope of the southeastern Arabian Sea contains a high %Corg (Singh et al., 2021). This morphogroup is dominated by the genera Bulimina, Uvigerina, Fursenkoina, Globocassidulina, and Reophax. These genera have been well documented in the high productivity areas of the world ocean, such as the eastern Pacific and the Arabian Sea (Schumacher et al., 2007; Uchimura et al., 2017). These genera are also primarily restricted to the upper sediment column and are classified as shallow infaunal (Jorissen et al., 1995; Kaithwar et al., 2020). As a result, the abundant presence of tapered and cylindrical benthic foraminifera morphogroup in a region should be used as an indicator of high organic carbon or primary productivity. Furthermore, the high organic matter preference of these morphogroups is further confirmed by their increased abundance during the times of high productivity such as the Bolling-Allerod and early Holocene (Verma et al., 2018). Besides, this morphogroup also had a negative correlation with bottom water dissolved oxygen, indicating that these genera thrive in regions where oxygen concentrations are depleted by the organic matter degradation and benthos respiration.

The flattened-ovoid morphogroup, like the tapered and cylindrical morphogroups, had similar environmental preference. A majority of the species belonging to the flattened-ovoid, tapered and cylindrical morphogroups are infaunal (Corliss and Fois, 1990). The flattened ovoid forms are abundant on the continental shelf and upper slope (top 1000 m) where dissolved oxygen concentration is at its lowest, as indicated by CCA (Figure 14; Figure 15). A similar low dissolved oxygen preference

of flattened ovoid forms was also suggested in the Gulf of Mexico and the Norwegian Sea (Corliss and Chen, 1988; Corliss and Fois, 1990), as well as in the eastern Arabian Sea, where its abundance declined during the times of weaker OMZs, such as the Heinrich Stadial 1 (HS1; Verma et al., 2018). The infaunal benthic foraminifera have a negative relationship with dissolved oxygen and a positive relationship with  $%C_{org}$ . In the Arabian Sea and other parts of the world ocean, infaunal benthic foraminifera are widely used as an indicator of the past OMZ variability (Mazumder and Nigam, 2014; Naik et al., 2017), and this work on living benthic foraminifera reconfirms it.

The epifaunal habitat of the biconvex morphogroup is well established. This group includes the genera *Cancris, Eponides, Bucella*, and *Hoeglundina*, which are usually grouped together under the epifaunal morphogroup (Nigam and Khare, 1999; Manasa et al., 2016). In the southeastern Arabian Sea, epifaunal forms were abundant at very shallow depths and on the lower slope, where organic carbon content is low. This finding is also supported by CCA, indicating a negative correlation between %C<sub>org</sub> and Biconvex morphogroup (Figure 14; Figure 15). Furthermore, their increased abundance was reported from both the shallow waters in the Arabian Sea (Nigam and Khare, 1999) as well as the deep water in the Norwegian Sea and Gulf of Mexico (Corliss and Chen, 1988; Corliss and Fois, 1990). The biconvex forms were also more abundant during the HS1 and HS2 (Verma et al., 2018), when surface primary productivity was lower. Biconvex morphogroup also showed a positive correlation with the bottom water dissolved oxygen, indicating that it thrives in well-oxygenated bottom water, such as shallow and deep-water. Planoconvex epifaunal benthic foraminifera showed the same positive/negative relationship with ambient environmental parameters. This group primarily includes the genera *Cibicidoides, Epistominella, Hanzawia*, and *Planulina*. Their



**Figure 15:** The representative species of the different morphogroups in the southeastern Arabian Sea. Tapered and Cylindrical (1. *Saccorhiza ramosa*; 2a-b. *Siphotextularia rolshauseni*; 3. *Bulimina aculeata*; 4. *Uvigerina multicostata*), Flattened-tapered (5a-c. *Bolivina obscuranta*), Spherical (6. *Lagena perculiaris*; 7. *Anturina haynesi*; 8. *Lagenosolenia eucerviculata*), Rounded-planispiral (9a-c. *Melonis pompilioides*), Biconvex (10a-c. *Cancris sagra*; 11a-c. *Buccella differens*), Planoconvex (12a-c. *Cibicidoides wuellerstrofi*; 13a-c *Hanzawaia concentrica*), Flattened-ovoid (14a-b. *Lenticulina lucidiformis*; 15a-c. *Cassidulina carinata*), Milioline (16. *Quinqueloculina parkeri*), Rounded-trochospiral (17a-c. *Haplophragmoides subglosum*; 18a-c. *Gyroidina quinqueloba*). Scale = 100 µm.

surface distribution revealed a high abundance in deep-water, where dissolved oxygen is abundant. This is also supported by CCA, which suggests a positive relationship between dissolved oxygen and the planoconvex group (Figure 14; Figure 15). The abundance and pore size of *Cibicidoides* is a well-established proxy for the bottom water oxygenation (Rathburn et al., 2018), but genera such as *Epistominella* are known as productivity indicators (Saraswat et al., 2005). This demonstrates that different genera/species within the same morphogroup may have distinct microhabitat preferences. Furthermore, planoconvex forms have been widely reported from the deep water in the world ocean (Corliss and Chen, 1988; Corliss and Fois, 1990), and their abundance was high during north Atlantic cold events such as Heinrich Stadial 1 and 2, when bottom water dissolved oxygen concentration was high due to lower primary productivity in the eastern Arabian Sea (Verma et al., 2018).

From the CCA plot, we report a positive relationship between flattened-tapered morphogroup and bottom water salinity as well as temperature. The relative abundance of this morphogroup was negatively correlated with  $C_{org}/N$  and water depth (Figure 14; Figure 15). This is quite intriguing, because flattened-tapered genera, such as *Bolivina*, were previously thought to be OMZ indicator species. It was more abundant between 0-250 m and 1000-1500 m water depths, where  $C_{org}$  concentration was comparatively low. Similarly, during HS1, when primary productivity was low, an increased percentage of flattened-tapered foraminifera was reported from the eastern Arabian Sea (Verma et al., 2018). A similar preference was reported from the Norwegian Sea, where it was found in abundance in shallow waters with low organic carbon. All these justify its negative relationship with  $C_{org}/N$ , such as Heinrich Stadials when the contribution of land derived organic matter decreased. This summarises that the morphogroup prefers low organic carbon.

The spherical morphogroup is associated with shallow infaunal microhabitats, and its comparatively high abundance in the lower slope region suggests its preference for plenty of food availability as well as surplus dissolved oxygen. This habitat preference is confirmed by its positive correlation with  $C_{org}/N$  and water depth in CCA (Figure 14; Figure 15). *Lagena, Baggina, Oolina,* and *Fissurina* are the major contributors to this morphogroup and all these genera are frequently found near the high riverine influx areas such as in front of the Ganga Brahmaputra rivers (Manasa et al., 2016). The southeastern Arabian Sea is a high marine organic carbon zone, and the relative abundance of this morphogroup is lower than that of others. Other studies have confirmed its infaunal microhabitat and preference for low levels of marine organic carbon (Corliss and Chen 1988; Corliss and Fois, 1990). Furthermore, Verma et al. (2018) confirmed its high abundance during the last glacial maximum, when the northeast monsoon was dominant, resulting in a high  $C_{org}/N$  in the study area. This morphogroup has a negative relationship with bottom water salinity and temperature, which

corresponds well with the regional hydrography and can be used to interpret the area's past environmental conditions. The rounded-trochospiral and rounded-planispiral morphogroups exhibit similar environmental preferences (Figure 14; Figure 15). This simply means that these morphogroups prefer food from the land. The rounded-trochospiral are usually epifaunal oxygen-loving species that increase in abundance with water depth, particularly after OMZ (> 1500 m). The same is true for the Gulf of Mexico, where its abundance increased abruptly after 1000 m water depth (Corliss and Fois, 1990). The most common genera in this morphogroup from the southeastern Arabian Sea are Gyroidina and Haplophragmoides. The past records confirm their preference for land-derived food, as their abundance decreased during the periods of reduced rainfall, such as Heinrich stadials 1 and 2. The rounded-planispiral forms are commonly thought of as an infaunal benthic group, with abundance increasing from southeast to northwest or from the Bay of Bengal side to the Arabian Sea side. The Bay of Bengal water contains a large amount of land-derived nutrients as well as organic matter, as evident from an increase in the %Corg/N. CCA results also revealed a positive correlation between %Corg/N and rounded-planispiral morphogroup, indicating that the land-derived food is preferred by genera such as Nonion, Nonionella, Melonis, and Pullenia. Its abundance also decreased during the periods of weaker southwest monsoon, such as HS1 and HS2.

In the southeastern Arabian Sea, two major-morphogroups, epifaunal and infaunal benthic foraminifera have a positive/negative relationship with %C<sub>org</sub> and dissolved oxygen. This indicates its applicability in the global ocean. However, a few workers prefer to use the terms rounded symmetrical benthic foraminifera (RSBF) and angular asymmetrical benthic foraminifera (AABF) over Epifaunal and Infaunal, respectively. In this work, we also checked that the factors affecting the relative abundance of the RSBF and AABF (Supplementary Figure 1). CCA of RSBF, AABF, and other morphogroups with the ambient physico-chemical parameters, reveals that AABF and RSBF do not show a positive/negative relationship with %C<sub>org</sub> and dissolved oxygen. This could be due to the fact that infuanal morphogroups such as rounded-planispiral, spherical, and flattened-ovoid are frequently considered under RSBF, not AABF (Manasa et al., 2016; Suokhrie et al., 2018). We recommend that workers should avoid using AABF and RSBF and instead focus on other morphogroups.

#### Conclusions

The relationship between living benthic foraminifera morphogroups and environmental parameters was tested in the southeastern Arabian Sea. We demonstrate a definite distribution pattern of a majority of the morphgroups in the surface sediment of the studied region. We conclude that infaunal morphogroups such as tapered/cylindrical and flattened ovoid are found in oxygen deficient zone which is caused by the high organic carbon degradation. Additionally, epifunal morphogroups such as

biconvex and planoconvex represent well-ventilated bottom water conditions. Besides this, other morphogroups, namely, flattened-tapered, spherical, rounded-trochospiral and rounded-planispiral forms were abundant at deep-water stations where land derived organic matter was abundant (as suggested by the high  $%C_{org}/N$ ) with low bottom water salinity and temperature. The environmental preferences of the living benthic foraminifera morphogroups delineated from the southeastern Arabian Sea, can be used to reconstruct past environmental conditions.

#### Acknowledgements

Authors are thankful to the Director, CSIR-National Institute of Oceanography, Goa, India for the permission to publish the results. DPS thanks the Indian Institute of Technology Roorkee for providing basic infrastructure for the completion of this work. RS is thankful to the Department of Science and Technology [DST/CCP/NCC&CV/130/2017(G)] and the Council of Scientific and Industrial Research, India for the financial support under the Young Scientist Research Scheme. Authors thank Dr. C.P. Babu for the help in organic carbon and nitrogen analysis. We also thank Professor Michael A. Kaminski, Department of Geosciences, College of Petroleum Engineering and Geosciences, King Fahd University of Petroleum and Minerals, Kingdom of Saudi Arabia, and one anonymous reviewer for taking the time and effort to review our work.

Author contribution: DPS conceived the idea and further developed with RS and RP. DPS has done the benthic foraminifera picking, identification and wrote the original draft. RS has acquired the funding and edited and reviewed the manuscript. The manuscript has been read and approved by all named authors.

**Funding:** The financial support is received from Department of Science and Technology [DST/CCP/NCC&CV/130/2017(G)].

#### **Declarations**

Ethics approval and consent to participate: Not applicable.

Consent for publication: Yes.

Competing interests: The authors declare no competing interests.

Availability of data and materials: The dataset is available in Supplementary Table 1.

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 Table 1: The living benthic foraminifera species belonging to different morphogroups in the southeastern Arabian Sea.

Tapered & Cylindrical	Biconvex	Rounded-trochospiral
Bathysiphon discreta	Cancris auriculus	Gyroidinoides soldanii
Bathysiphon folini	Cancris cf. penangensis	Gyroidina pilasensis
Saccorhiza ramosa	Cancris penangensis	Gyroidina tenera
Reophax brevis	Cancris sagra	Gyroidina cf. guadalupensis
Reophax rostrata	Eponides umbonatus	Gyroidina pilasensis
Trochammina conica	Osangularia bengalensis	Gyroidina quinqueloba
Gaudryina baccata	Buccella tenerrima	Gyroidina io
Eggerella humboldti	Buccella differens	Cribrostomoides nitida
Eggerelloides scaber	Pararotalia sp.	Haplophragmoides canariensis
Siphotextularia masudai	Rotalidium annectens	Haplophragmoides evolutum
Pseudogaudryina		
triangulate	Operculina inaequilaterais	Haplophragmoides sphaeriloculus
Nodosaria brevis	Rotorbinella bikinensis	Haplophragmoides subglobosum
Amphicoryna bilocularis	Cancris sp.	Haplophragmoides symmetricus
Hyalinonetrion elongata	Gyroidina cf. neosoldanii	Haplophragmoides bradyi
Globocassidulina porrecta	Hoeglundina heterolucida	Cystammina sp.
Globocassidulina		Ammoglobigerina
subglobosa		globigeriniformis
Ehrenbergina pacifica	Planoconvex	Portatrochammina eltaninae
Hopkinsina atlantica	Neoconorbina terquemi	
Hopkinsinella glabra	Rosalina columbiensis	Rounded-planispiral
Stainforthia loeblichi	Cibicidoides mundula	Ammobaculites aff. exiguus
	$C^{1} \cdot \cdot 1 \cdot 1  11  C^{1}$	
Bulimina arabiensis	Cibicidoides wuellerstorfi	Eratidus foliaceus
Bulimina arabiensis Fursenkoina spinosa	Epistominella pulchella	Nonion glabrella
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata	Epistominella pulchella Epistominella exigua	Nonion glabrella Nonion granosum
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis	Epistominella pulchella Epistominella exigua Epistominella umbonifera	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata	Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp.	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata	Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis	<i>Epistominella pulchella</i> <i>Epistominella exigua</i> <i>Epistominella umbonifera</i> <i>Epistominella sp.</i> <i>Psuedoeponides equatoriana</i> <i>Amphistegina gibbosa</i>	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides	<i>Epistominella pulchella</i> <i>Epistominella exigua</i> <i>Epistominella umbonifera</i> <i>Epistominella sp.</i> <i>Psuedoeponides equatoriana</i> <i>Amphistegina gibbosa</i> <i>Hanzawaia concentrica</i>	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp.
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina porrecta	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina puedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina porrecta Uvigerina auberiana	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina porrecta Uvigerina auberiana Uvigerina af. asperula	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina porrecta Uvigerina auberiana Uvigerina af. asperula Uvigerina af. canariensis	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida Flattened-ovoid	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis Valvulineria hamanakoensis
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina aporrecta Uvigerina auberiana Uvigerina af. asperula Uvigerina af. canariensis Uvigerina longa	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida Flattened-ovoid Lenticulina calcaesfera	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis Valvulineria hamanakoensis Valvulineria minuta
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina aporrecta Uvigerina auberiana Uvigerina af. asperula Uvigerina af. canariensis Uvigerina longa Uvigerina aff. mediterranea	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida Flattened-ovoid Lenticulina calcaesfera Lenticulina crassa	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis Valvulineria hamanakoensis Valvulineria minuta Ammodiscus gullmarensis
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina pupoides Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina porrecta Uvigerina auberiana Uvigerina af. asperula Uvigerina af. canariensis Uvigerina longa Uvigerina af. mediterranea Uvigerina af. asperula	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida Flattened-ovoid Lenticulina calcaesfera Lenticulina lucidiformis	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis Valvulineria hamanakoensis Valvulineria minuta Ammodiscus gullmarensis Hyalinea balthica
Bulimina arabiensis Fursenkoina spinosa Bulimina aculeata Bulimina alazanensis Bulimina marginata Bulimina marginospinata Bulimina marginospinata Bulimina psuedoaffinis Bulimina pupoides Bulimina striata Neouvigerina ampullacea Neouvigerina ampullacea Veigerina adberiana Uvigerina af. asperula Uvigerina af. canariensis Uvigerina af. mediterranea Uvigerina af. asperula Uvigerina af. asperula	Cibicidoides wuellerstorfi Epistominella pulchella Epistominella exigua Epistominella umbonifera Epistominella sp. Psuedoeponides equatoriana Amphistegina gibbosa Hanzawaia concentrica Planulina foveolatiformis Tritaxis fusca Trochammina boltovskoyi Rosalina Candida Flattened-ovoid Lenticulina calcaesfera Lenticulina lucidiformis Lenticulina pliocaena	Eratidus foliaceus Nonion glabrella Nonion granosum Nonionella simplex Nonionellina labradorica Melonis cf. pompilioides Melonis cf. chathamensis Melonis affinis Melonis sp. Pullenia bulloides Pullenia salisburyi Nonionella aff. tumidela Ammomarginulina troptunesis Valvulineria hamanakoensis Valvulineria minuta Ammodiscus gullmarensis Hyalinea balthica

Uvigerina mediterranea	Neolenticulina peregrina
Uvigerina multicostata	Neolenticulina antarctica
Uvigerina peregrina	Allomorphina pacifica
Reussella aequa	Cassidulina aff. minuta
Reussella lavelaensis	Cassidulina angulosa
Fursenkoina cornuta	Cassidulina bradvi
Fursenkoina obliaua	Cassidulina carinata
Robertinoides oceanica	Cassidulina laevigata
Reussella sp.	
Fusenkoina rotundata	Flattened-tanered
Ruliminella evilis	Spiroplectamming hiformis
Globobuliming pacifica	Spirotextularia cf fistulosa
Astacolus insolitus	Boliving acquiis
Astacotas insolitas	Boliving advena
Sabariaal	Bolivina davena Bolivina obvodki
Spherical	Bolivina churchi
Aaercotryma glomeratum	Bolivina cincta
Baggina diversa	Bolivina compacta
Saccammina huanghaiensis	Bolivina currai
Lagena cf. flatulenta	Bolivina dilatata
Lagena perculiaris	Bolivina globulosa
Pygmaeoseistron hispidula	Bolivina inflata
Anturina haynesi	Bolivina jacksonensis
Oolina cf. globosa Fissurina aligera	Bolivina lowmani
caudimarginata	Bolivina mantaensis
Fissurina cassiparosa	Bolivina obscuranta
Lagenosolenia	
eucerviculata	Bolivina pacifica
Lagenosolenia neocincta	Bolivina mera
Lagenammina longicolli	Bolivina robusta
Hormosinella distans	Bolivina seminuda
Rotaliatinopsis	
semiinvoluta	Bolivina sp. A
Laticarinina Carinata	Bolivina spathulata
	Bolivina spinescens
	Bolivina striatula
	Bolivina victoriana
	Bolivina zanzibarica

**Supplementary Figure 1:** Canonical Correspondence Analysis plot between different benthic foraminifera morphogroups (including AABF and RSBF) and ambient environmental parameters.



**Table 1:** The Table contains sample location in the study area, ecological parameters, and relative abundance of various morphogroups along with absolute abundance of living benthic foraminifera.

Locations	Longitude (°E)	Latitude (°N)	Water Depth (m)	Temp (°C)	Salinity (psu)	DO (ml/l)	%Corg	%Corg/TN	Tapered & Cylindrical (%)	Spherical (%)	Biconvex (%)	Planoconvex (%)	Flattened ovoid (%)	Flattened- tapered (%)	Rounded- trochospiral (%)	Rounded – planispiral (%)	Epifaunal (%)	Infaunal (%)	Absolute Abundance (#/g sed)
G2	77.3513	8.0343	25	27.45	35.20	4.25	0.82		28.44	11.01	7.34	17.43	0.00	15.60	18.35	1.83	43.12	56.88	4
MC09	77.9493	8.0778	26	27.42	35.20	4.21	4.30		24.71	2.65	4.41	4.41	4.12	40.00	7.65	12.06	16.47	83.53	26
MC25	76.9163	8.3365	46	26.32	35.12	3.45	0.71	6.22	23.40	2.48	4.26	30.14	1.06	29.08	8.51	1.06	42.91	57.09	15
MC10	77.9551	7.7329	50	26.08	35.09	3.3	1.64		31.94	14.19	5.16	8.39	1.94	15.16	14.19	9.03	27.74	72.26	41
G1	77.3521	7.8649	50	26.08	35.09	3.3	3.06		32.52	14.72	11.04	8.90	3.68	4.91	8.28	15.95	28.22	71.78	13
MC08	78.7533	8.9201	58	25.96	35.19	3.39	1.33	12.80	24.46	0.00	0.00	0.66	5.45	64.96	3.80	0.66	4.46	95.54	15
G3	76.7964	8.246	60	25.03	35.11	2.82	2.26		41.18	0.00	0.00	0.00	2.94	52.94	0.00	2.94	0.00	100.00	31
G4	76.7007	8.1736	101	20.66	35.12	1.13	1.03		8.33	0.00	29.17	20.83	0.00	25.00	4.17	12.50	54.17	45.83	39
MC23	77.2149	7.4062	107	20.15	35.10	0.99	3.94		42.86	0.00	14.29	14.29	0.00	28.57	0.00	0.00	28.57	71.43	16
MC11	77.9495	7.5837	110	19.90	35.08	0.93	2.41		14.29	0.00	11.43	37.14	0.00	37.14	0.00	0.00	48.57	51.43	16
MC07	78.7529	8.8929	152	16.28	35.03	0.53	2.46	8.96	27.27	0.00	9.09	9.09	0.00	54.55	0.00	0.00	18.18	81.82	13
MC06	78.7429	8.8648	215	13.61	35.07	0.69	3.22	9.50	25.00	0.00	0.00	0.00	25.00	50.00	0.00	0.00	0.00	100.00	448
MC12	77.9501	7.5372	225	13.61	35.05	0.53	6.20		40.34	7.56	1.68	13.45	13.45	10.08	2.52	10.92	17.65	82.35	22
G5	76.636	8.139	260	12.74	35.06	0.52	2.83		23.14	1.65	14.88	6.34	31.40	6.34	6.61	9.64	27.82	72.18	17
MC26	76.6045	8.1228	503	10.02	35.05	0.35	7.24	8.90	16.73	6.05	29.18	16.01	7.12	4.98	19.57	0.36	64.77	35.23	232
MC05	78.7499	8.815	510	9.94	35.04	0.48	4.26	6.71	45.59	0.00	17.65	2.94	0.00	4.41	19.12	10.29	39.71	60.29	94
MC21	77.1256	7.1157	537	9.76	35.04	0.36	4.33	17.84	21.31	9.84	3.28	9.84	9.84	1.64	18.03	26.23	31.15	68.85	93
MC20	77.1333	7.1037	717	8.44	34.99	0.51	6.54	14.25	26.75	1.23	8.23	9.05	16.05	6.17	10.70	21.81	27.98	72.02	111
MC04	78.7295	8.7439	745	8.26	34.99	0.63	4.67	6.47	40.00	0.00	12.00	8.00	0.00	20.00	8.00	12.00	28.00	72.00	77
MC27	76.5647	8.0708	764	8.10	34.98	0.62	8.32	9.17	25.00	4.17	4.17	11.46	39.58	12.50	2.08	1.04	17.71	82.29	97
MC28	76.5091	8.019	970	6.78	34.97	0.96	8.27	9.79	24.39	0.00	17.07	9.76	19.51	14.63	12.20	2.44	39.02	60.98	42
MC03	78.7295	8.6324	1002	6.56	34.95	1.07	5.19	5.40	100.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	100.00	110
MC19	77.1316	7.078	1045	6.38	34.97	1.09	6.95	7.02	10.00	0.00	40.00	10.00	0.00	30.00	10.00	0.00	60.00	40.00	36
MC13	77.9512	7.3795	1100	6.14	34.97	1.18	5.10	10.13	78.07	0.53	2.14	1.60	0.00	12.83	1.60	3.21	5.35	94.65	161

MC29	76.3936	7.9379	1210	5.65	34.94	1.33	7.48	9.93	38.60	0.00	14.04	12.28	1.75	26.32	3.51	3.51	29.82	70.18	14
MC02	78.7365	8.4279	1250	5.50	34.92	1.43	5.39	5.47	53.33	8.33	1.67	3.33	8.33	25.00	0.00	0.00	5.00	95.00	80
MC18	77.1229	7.0533	1327	5.17	34.92	1.53	6.59	6.40	20.65	6.52	9.78	7.61	1.09	44.57	5.43	4.35	22.83	77.17	145
MC31	76.0441	7.6616	1454	4.49	34.88	1.78	2.24	10.00	59.38	3.13	9.38	9.38	3.13	0.00	15.63	0.00	34.38	65.63	3
MC30	76.2243	7.8288	1506	4.25	34.87	1.88	5.54	11.74	0.00	0.00	0.00	66.67	0.00	0.00	0.00	33.33	66.67	33.33	23
MC14	77.9514	7.31	1530	4.15	34.87	1.93	5.26	7.73	14.29	2.86	17.14	42.86	5.71	2.86	2.86	11.43	62.86	37.14	263
MC15	77.9458	7.2254	1540	4.10	34.87	1.95	6.93	8.89	30.77	0.00	0.00	53.85	0.00	0.00	0.00	15.38	53.85	46.15	81
MC01	78.7249	8.1081	1550	4.09	34.86	2.03	5.45	8.78	25.00	0.00	12.50	12.50	12.50	12.50	12.50	12.50	37.50	62.50	47
MC32	75.821	7.515	1704	3.57	34.82	2.31	3.26	11.72	27.27	0.00	18.18	9.09	0.00	9.09	18.18	18.18	45.45	54.55	13
MC17	77.1242	6.981	1802	3.25	34.81	2.48	5.48	12.80	16.28	3.49	0.00	39.53	0.00	8.14	23.26	3.49	62.79	31.40	36
MC60	79.0983	8.0191	1887	3.01	34.80	2.85	3.59		20.00	15.00	0.00	15.00	5.00	0.00	15.00	30.00	30.00	70.00	18
MC54	76.0121	5.9872	2065	2.68	34.78	2.86	1.35	14.35	19.30	9.65	4.68	12.87	11.11	9.06	25.15	7.31	42.69	56.43	6
MC16	76.9748	6.6911	2080	2.58	34.78	2.86	4.19	12.50	33.91	12.17	6.96	17.39	0.00	3.48	26.09	0.00	50.43	49.57	18
MC59	79.0058	7.8447	2080	2.62	34.78	3.09	5.15	8.83	81.82	3.03	0.00	3.03	0.00	3.03	9.09	0.00	12.12	87.88	15
MC55	77.0096	5.9974	2238	3.06	34.76	3.06	2.13	14.55	0.00	0.00	0.00	28.57	0.00	28.57	42.86	0.00	71.43	28.57	4
MC56	77.8429	5.9847	2460	2.09	34.75	3.25	1.96	4.83	100.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	100.00	23
MC53	75.0281	5.9906	2750	1.84	34.74	3.46	3.03	5.50	36.36	0.00	9.09	9.09	9.09	18.18	9.09	9.09	27.27	72.73	9
MC58	79.0225	7.2171	2750	2.06	34.76	3.48	5.87	7.18	11.11	0.00	33.33	0.00	33.33	22.22	0.00	0.00	33.33	66.67	149
MC57	78.9799	5.9864	2980	2.05	34.74	3.3	2.56	8.67	25.00	0.00	0.00	0.00	0.00	75.00	0.00	0.00	0.00	100.00	5